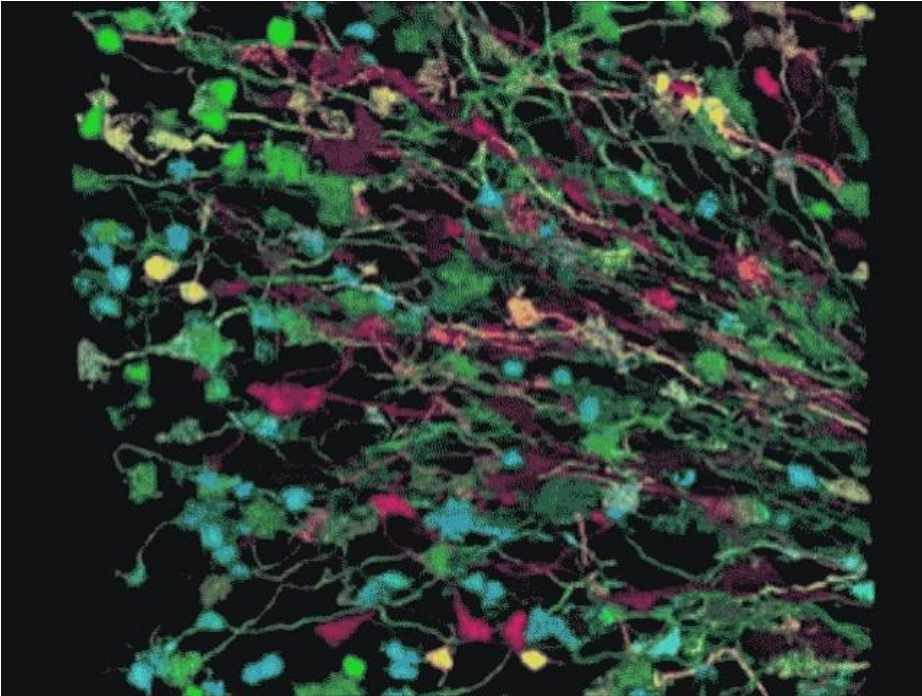


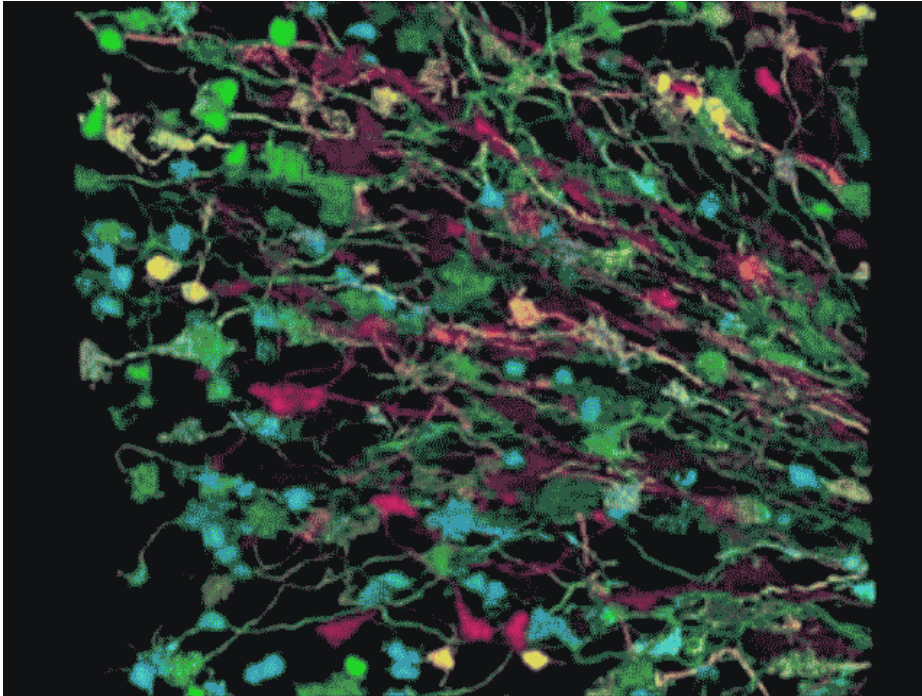
The brain: networks of neurons



Mouse cerebellum

Lichtman et al. 2008

The brain: networks of neurons



Mouse cerebellum

Lichtman et al. 2008

1 mm³

~1.5 year, 1 *petabyte* (10^{15})

Mouse brain:

~2000 years, 1 *exabyte* (10^{18})

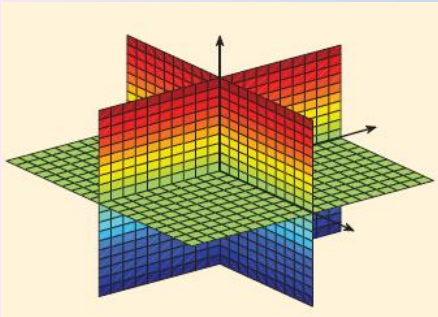
Human cortex:

~ 10^7 years, 10 *zettabytes* (10^{21})

World storage: 300 *exabytes*

~ on January 2015

Scalable Inference in Large Neural Systems



Allie Fletcher
Nov 5, 2015

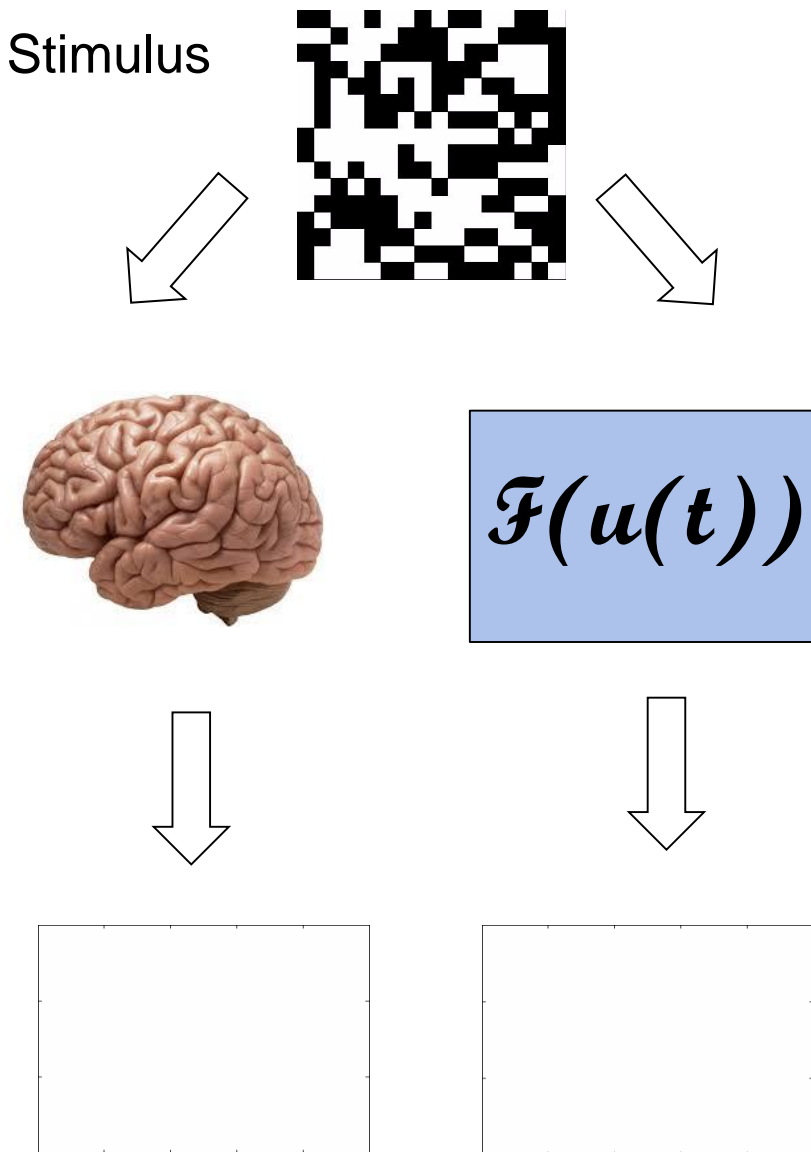


UC Santa Cruz Electrical Engineering
UC Berkeley Redwood Center, HWNI
UCLA Statistics, Math, & EE— Jan 2016

Outline

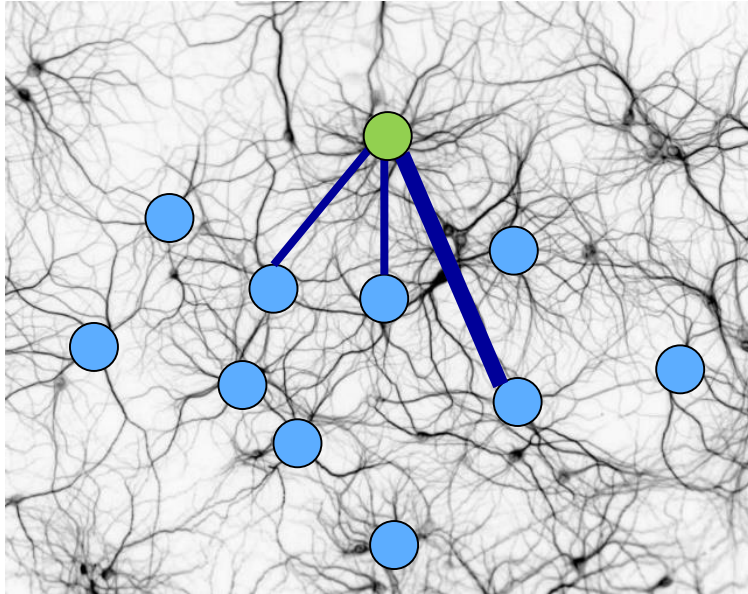
- Neural mapping via multineuron excitation
 - Background: Neuroscience
 - Optogenetic stimulation
 - Estimation via graphical models
- Network connectivity via Ca^{2+} imaging
 - Large scale but indirect and smoothed
 - Network model captures dynamics
 - Scalable accurate algorithm
- Receptive Field of Retinal Ganglion Cells
 - Spatio-temporal filtering
 - Improved identification with limited data

Neuroscience: Estimation Challenges



- Large-scale
 - ~86 billion neurons
 - V1 alone 140 million
- Sparse: ~1000 connections
- Complex:
 - Nonlinear dynamics
 - Feedback
- Indirect measurements
- Incomplete data
- Limited *in vivo* collection

I. Connectivity Detection to a Single Neuron



Problem:

Detect connections to one neuron

Goal:

Reduce trials and computation

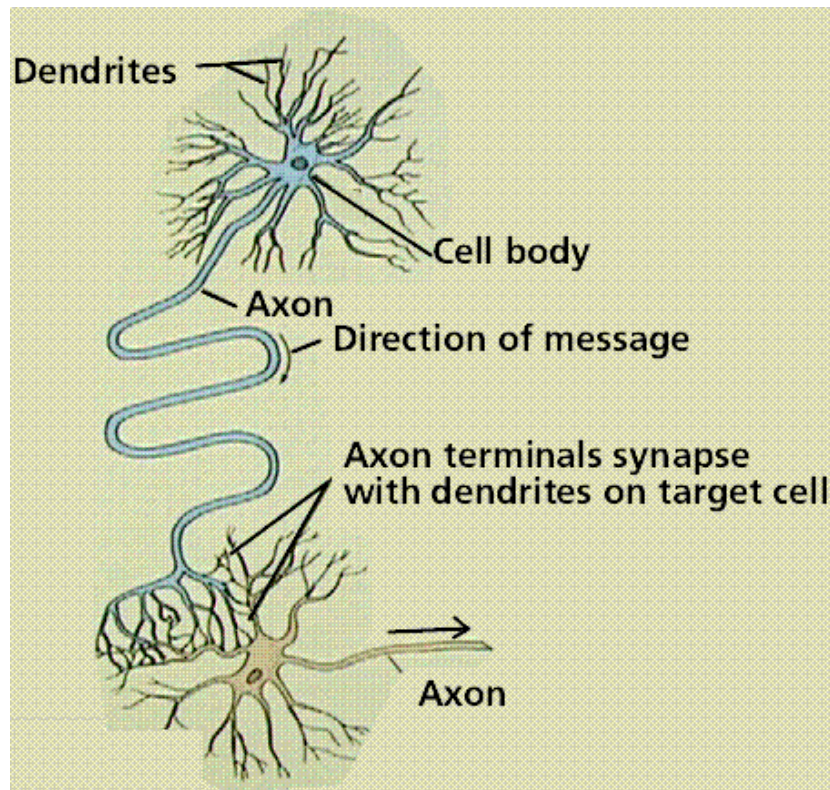
How:

Optogenetics

Improved “decoding” via subset stimulation

Neuron: Basic Anatomy

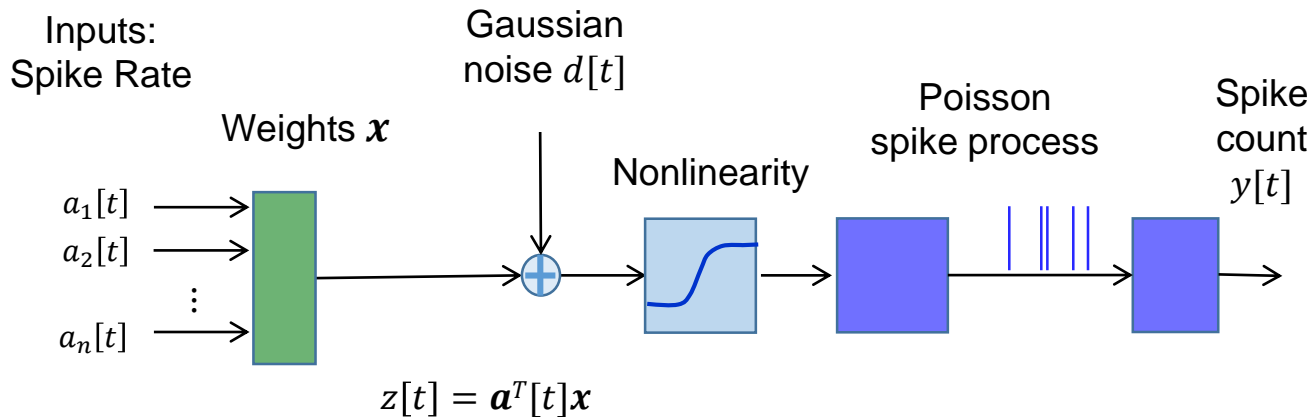
Neuron: Basic cell for information processing



■ Components

- **Dendrites:**
Filaments receive signals
- **Soma:** cell body
- **Axon:**
Outputs electrical signals to neurons or motor functions
- **Synapses:**
Junctions of axons and dendrites

Neuron: Linear-Nonlinear Poisson Model



$$z[t] = \sum_j x_j a_j[t]$$

df

$$\lambda[t] = f(z[t] + d[t])$$

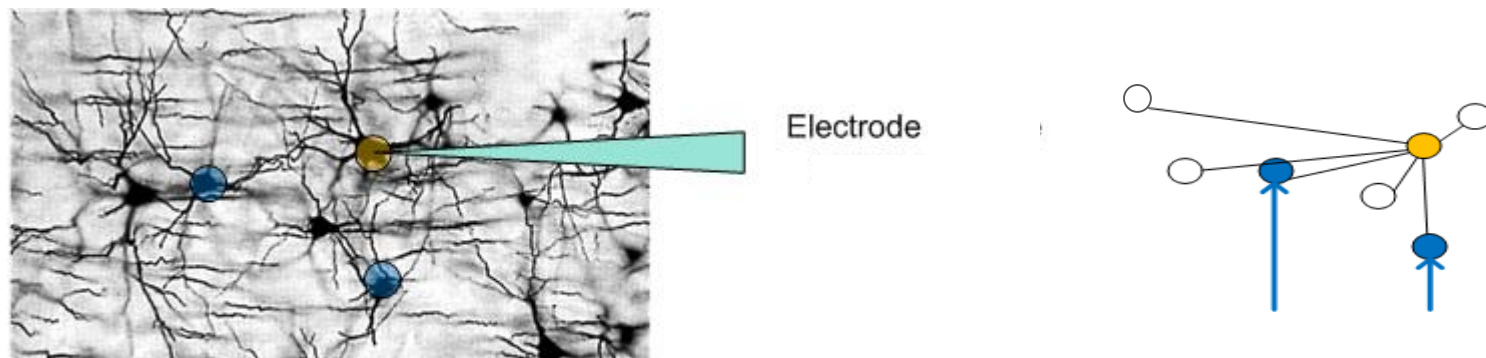
df

$$y[t] \sim \text{Poisson}(\lambda[t])$$

- **Functional Model: simple input-output**
 - Effectively captures average rates
 - Windows: 10 to 200 spikes/s
- **Three-stage LNP: Linear + Nonlinearity + Poisson process**
- **Connectivity: Identify weights x**
- **Biological models with feedback for precise timing later...**

Classic Connectivity Detection

Measure at post-synaptic neuron of interest



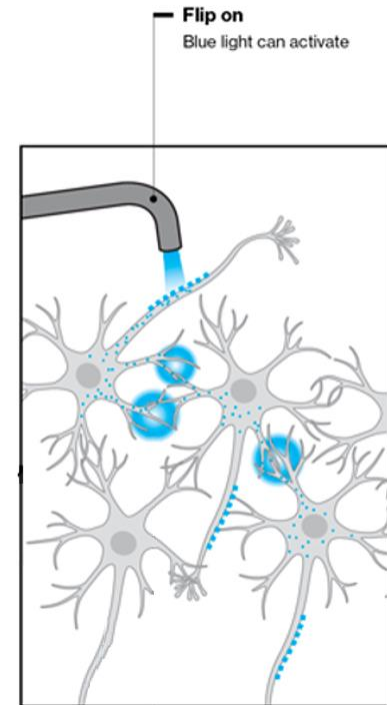
Stimulate potential pre-synaptic neurons: one at a time

- Problems: many measurements per test neuron
 - Most neurons are not connected
 - Noisy system with many exogenous inputs
 - Test neurons die
- Can we do better?

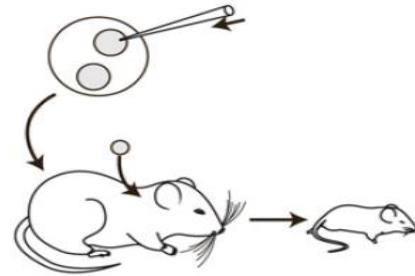
New Technologies: Optical Stimulation

Connectivity Detection:

- Genetically modified neuron
 - Photosensitive protein
- Optically activate test neurons
- Greater spatial precision
 - Pinpoint individual neurons
 - Multiple neurons at once

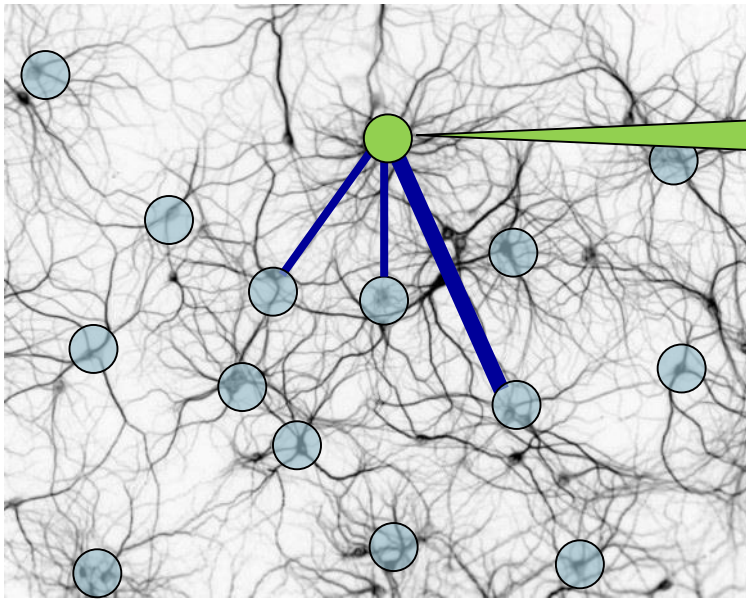


Wang, Hasan & Seung 2009





Transgenic mice

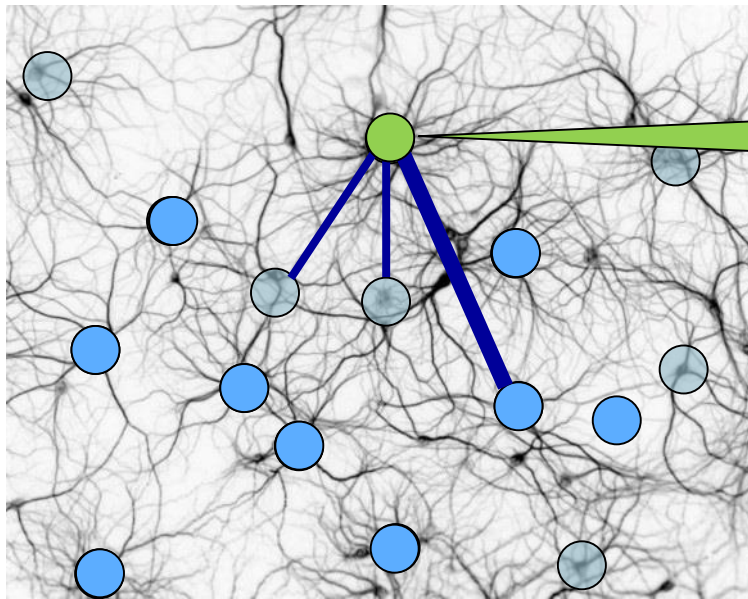
Connectivity via Multineuron Excitation



Spike recording
Average over time windows

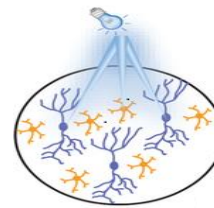
-  Potential presynaptic neurons
-  Synapses or connections

Detection via Multineuron Excitation



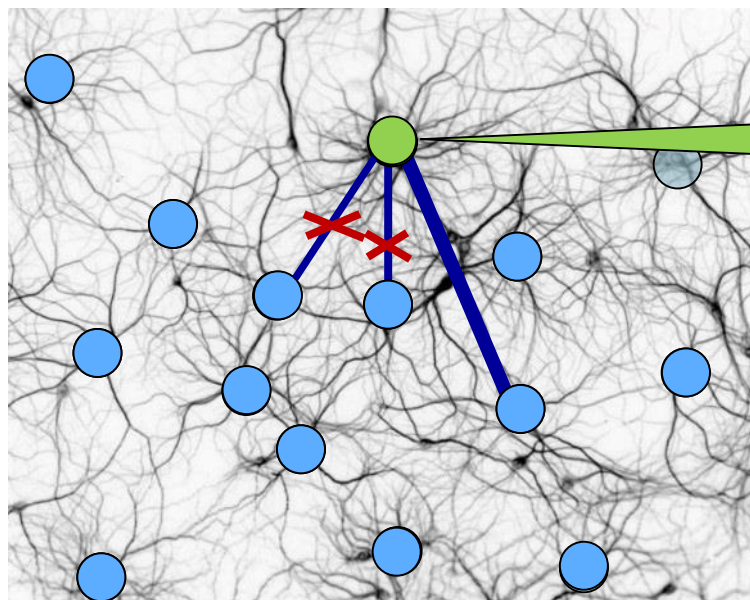
Strong response

Nonconnection



- Stimulate subsets of neurons at a time
 - Increase probability of response
 - Fewer wasted trials

Detection via Multi-Neuron Stimulation



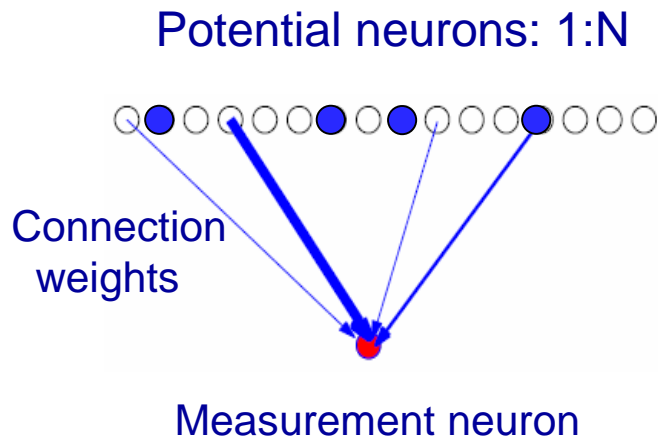
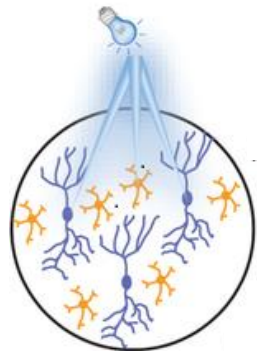
~~Low response~~
Low response

Sub-threshold
Sub-threshold

- Stimulate subsets of neurons at a time
 - Increase probability of a response
 - Less measurements wholly wasted
- Benefits
 - Weak sub-threshold connection
 - More reliable with less data

Hu & Chklovskii NIPS 2010,
Fletcher et al NIPS 2011

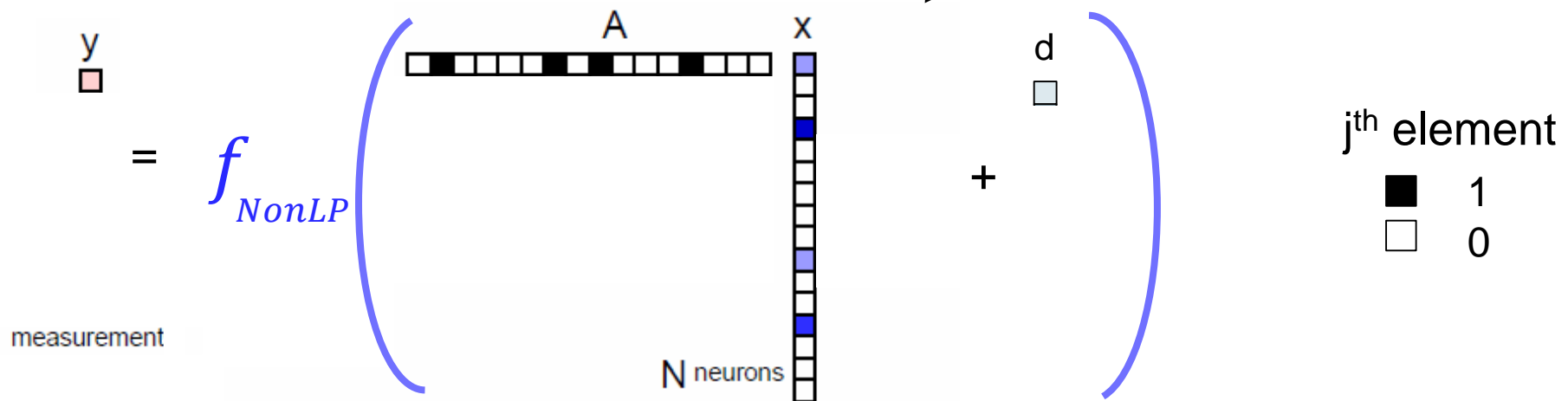
Multi-neuron Excitation: Model



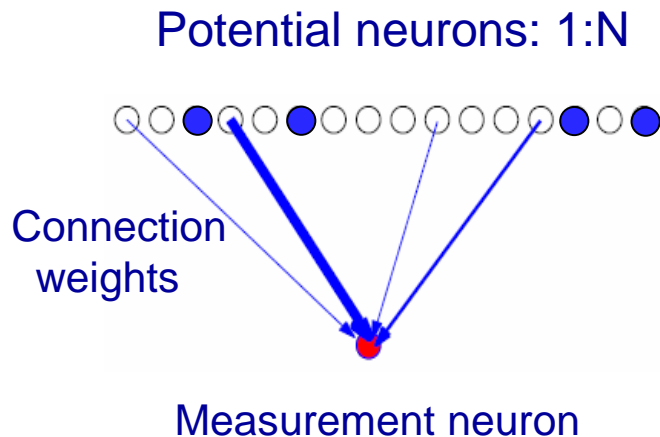
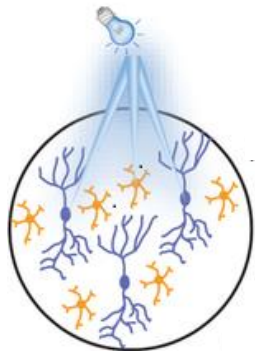
j^{th} neuron

- Excited
- Not excited

Excited: {2, 7, 9, 13}



Multi-neuron Excitation: Model

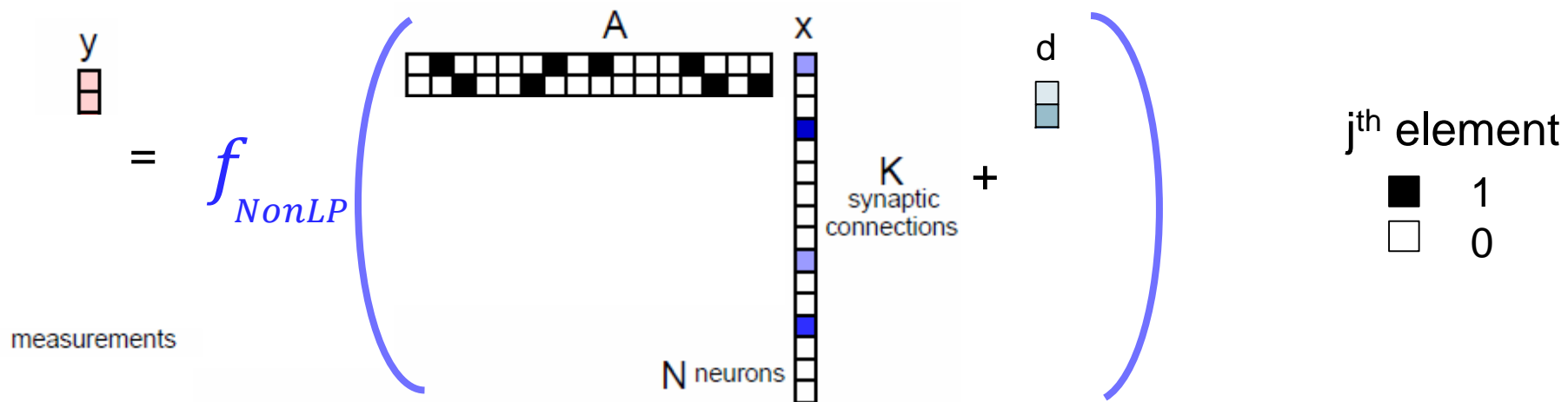


j^{th} neuron

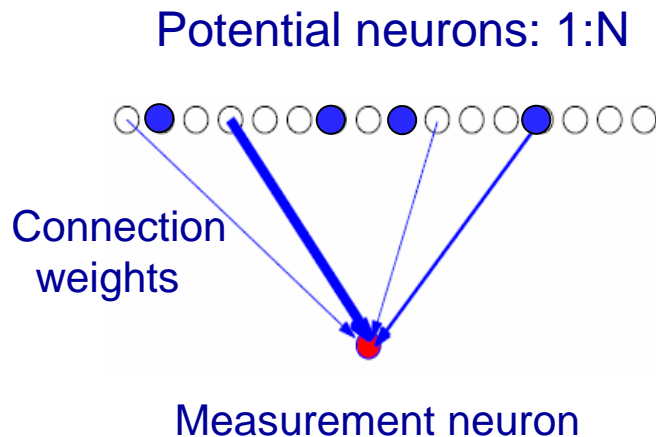
- Excited
- Not excited

Second trial
Different random subset
Excited: {3, 6, 14, 16}

"1" entries: {3, 6, 14, 16}



Multi-neuron Excitation: Model

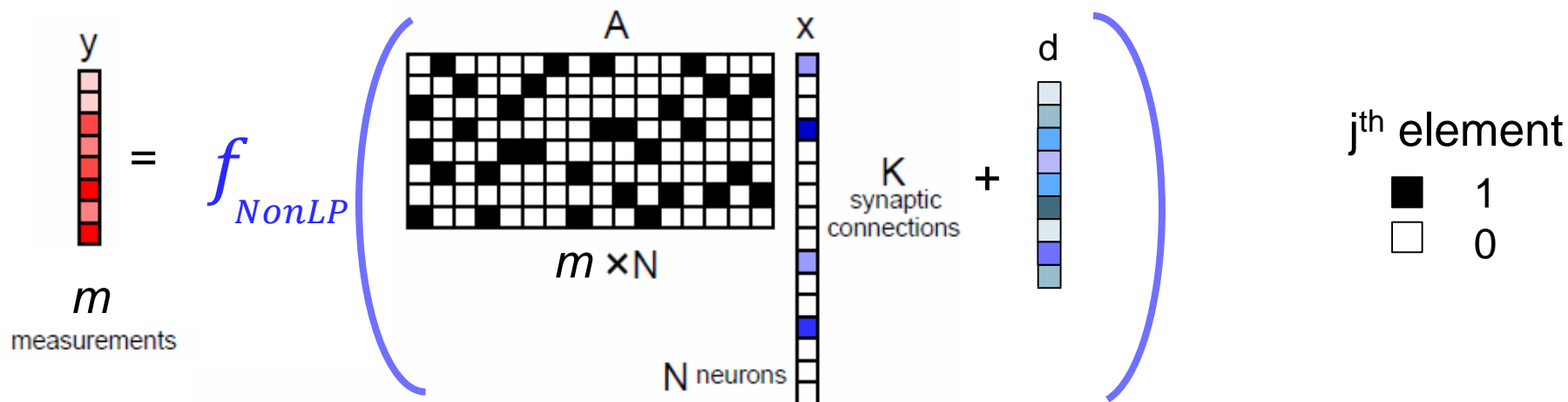


j^{th} neuron

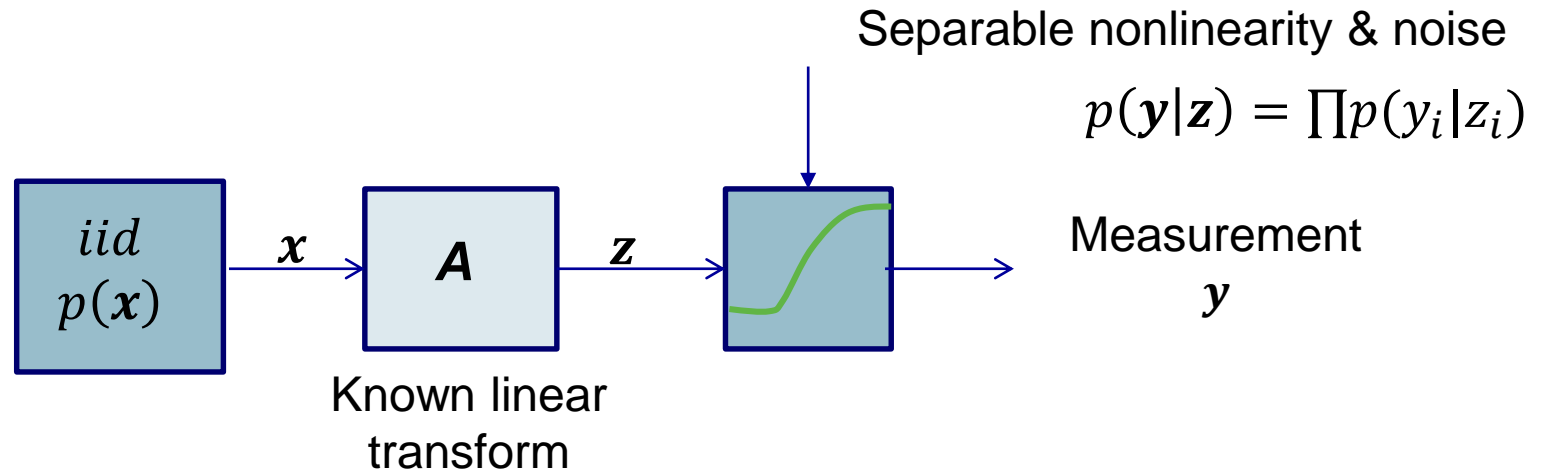
- Excited
- Not excited

m^{th} excitation

m trials or "measurements"



Bayesian Nonlinear Generalized Linear Models



- **Problem:** Estimate \mathbf{x} and \mathbf{z} given \mathbf{y} and \mathbf{A}
- **Bayesian formulation:** general system class
 - Prior $P_X(\mathbf{x})$ incorporates constraints, like sparsity
 - $P_{Y|Z}(\mathbf{y}|\mathbf{z})$ models output: nonlinearities, randomness
- **Challenge:** optimal estimation is hard
 - Components of vector \mathbf{x} are **coupled** in \mathbf{z}

Example: Sparse recovery

$$y = Ax + d$$

$p(x) = \text{Sparse prior}$
 $p(y|z) = \text{Gaussian}$

- **Problem:** Given \mathbf{A} and \mathbf{y} , recover sparse \mathbf{x}
- Many applications
 - Communication channels, linear inverse problems
 - Wavelet image reconstruction
 - Regularized linear regression, classification
 - Compressed sensing...
- Now many algorithms, theoretical analyses, ...

Divide & Conquer with Graphical Models

- Subdivide & conquer

$$p_{X|Y}(\mathbf{x}|\mathbf{y}) = \prod_{j=1}^n p(x_j) \prod_{i=1}^m p(y_i|\mathbf{x})$$

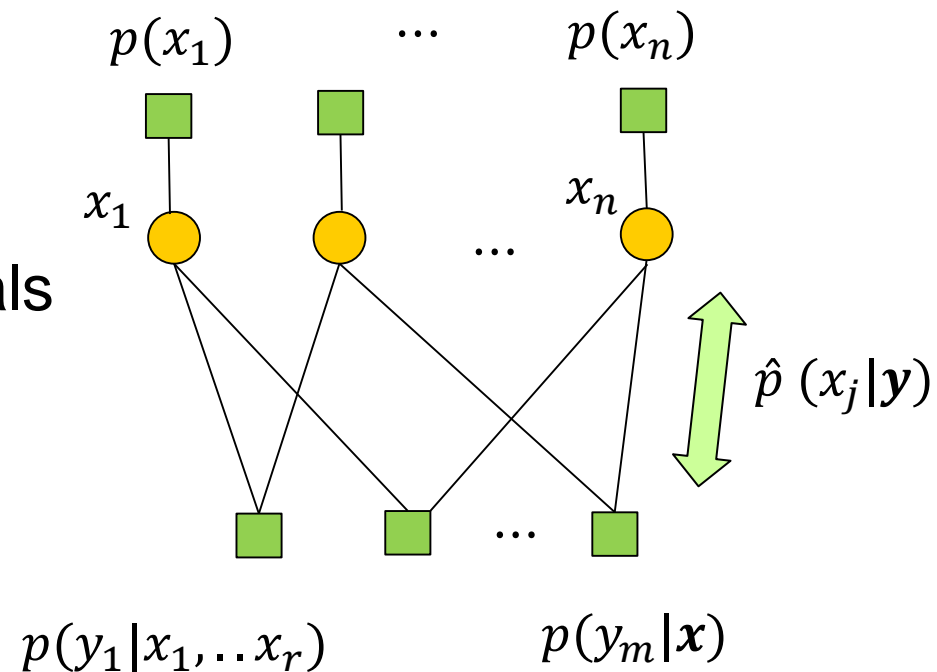
- Few “smaller” components

- Few variables
- Limited connections

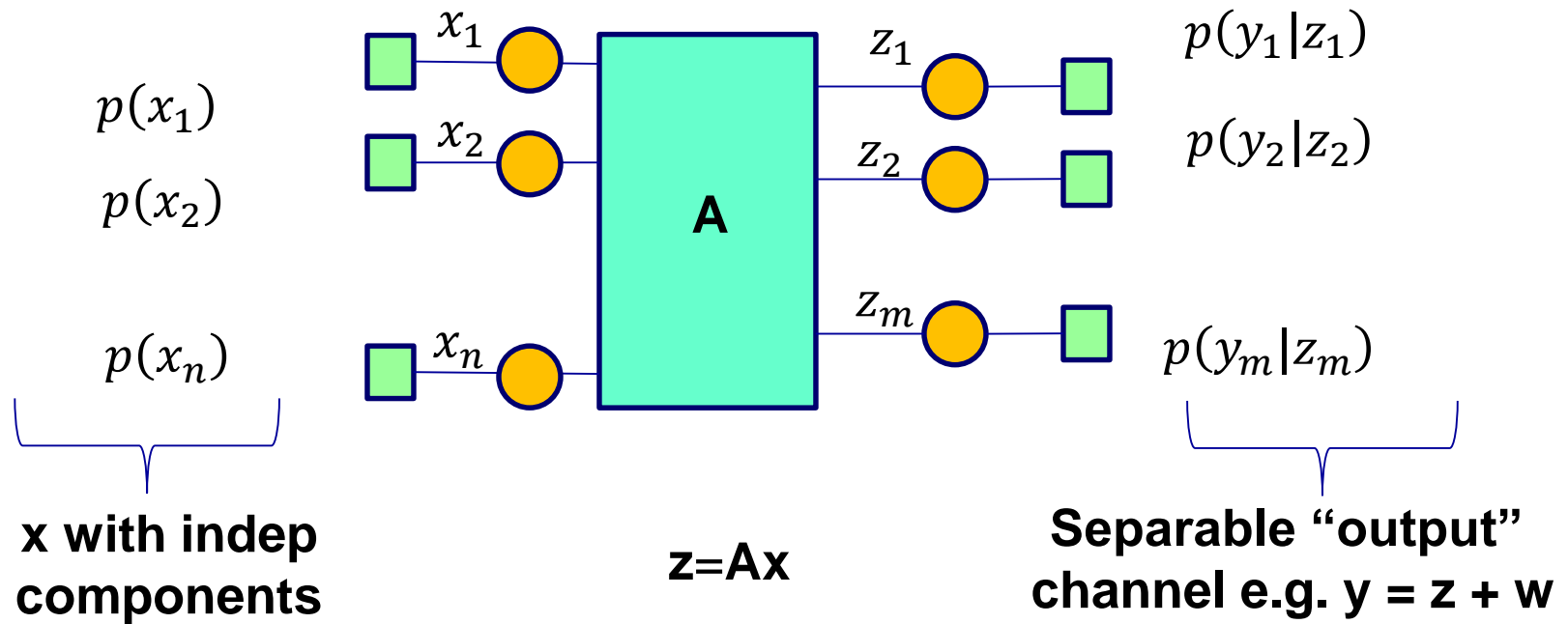
- Message passing:

- Iteratively update marginals
- Global estimation local

- But random A is dense!

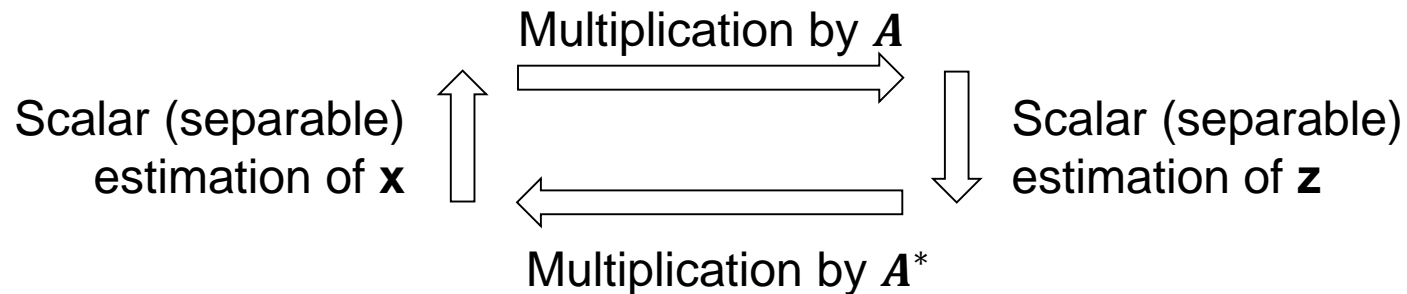
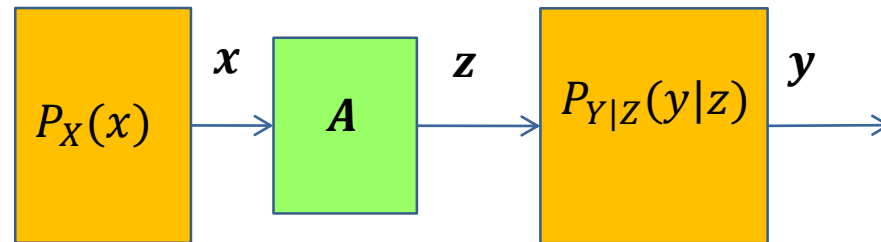


"Graphical Model" for GLM



- Assume separable priors & likelihoods
- The posterior density $p(x|y)$ factors into:
 - $m + n$ scalar terms; and
 - Linear constraint $z = Ax$

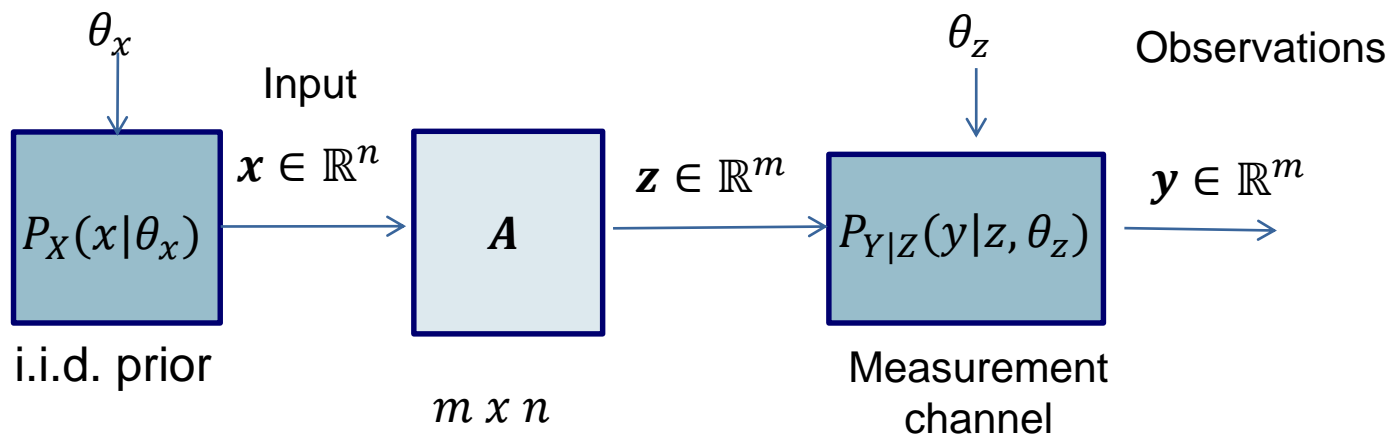
Generalized Approximate Message Passing (GAMP)



- Gaussian & quadratic approximations
- Asymptotic guarantees
- Low complexity: $O(mn)$ each iteration
- Classic AMP*: separable distributions & AWGN
- GAMP: KNOWN nonlinearities

AMP Donoho, Maleki, Montanari 09, Bayati & Montanari 10,
GAMP Rangan et al 10, HyGAMP Fletcher et al 11, ...

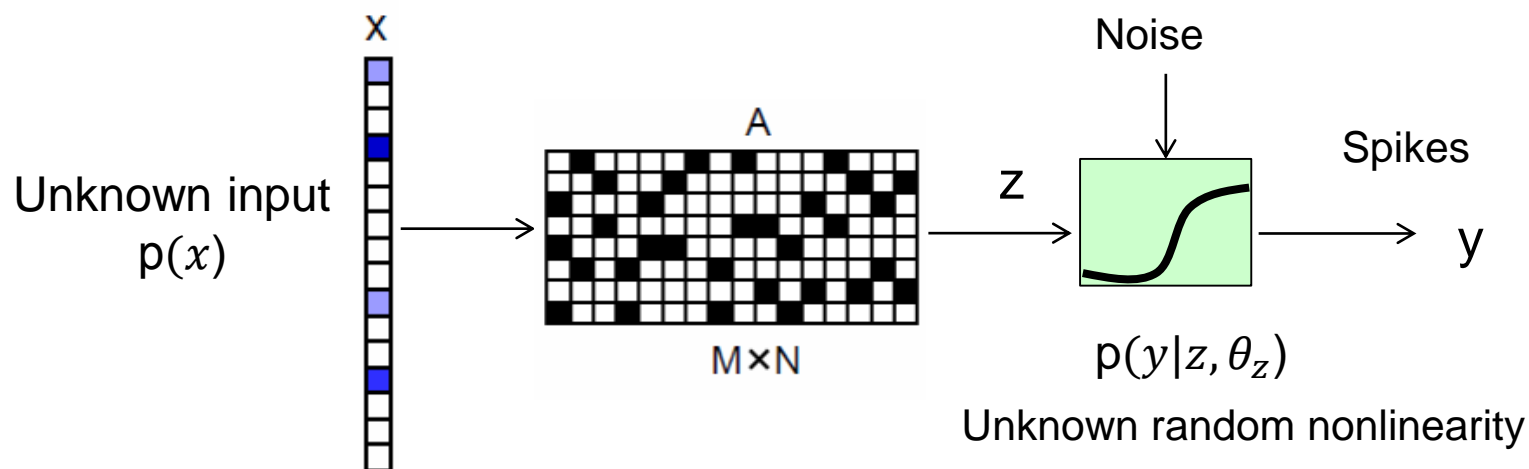
Theorem: Joint Estimation & Learning with Adaptive GAMP



- GLM with unknown parameters θ_x and θ_z
 - Unknown prior, nonlinearities, noise...
- Joint estimation learning problem: Given y and A :
 - Estimate input x and z ,
 - Learn parameters θ_x and θ_z in distribution
 - **Consistent estimator**



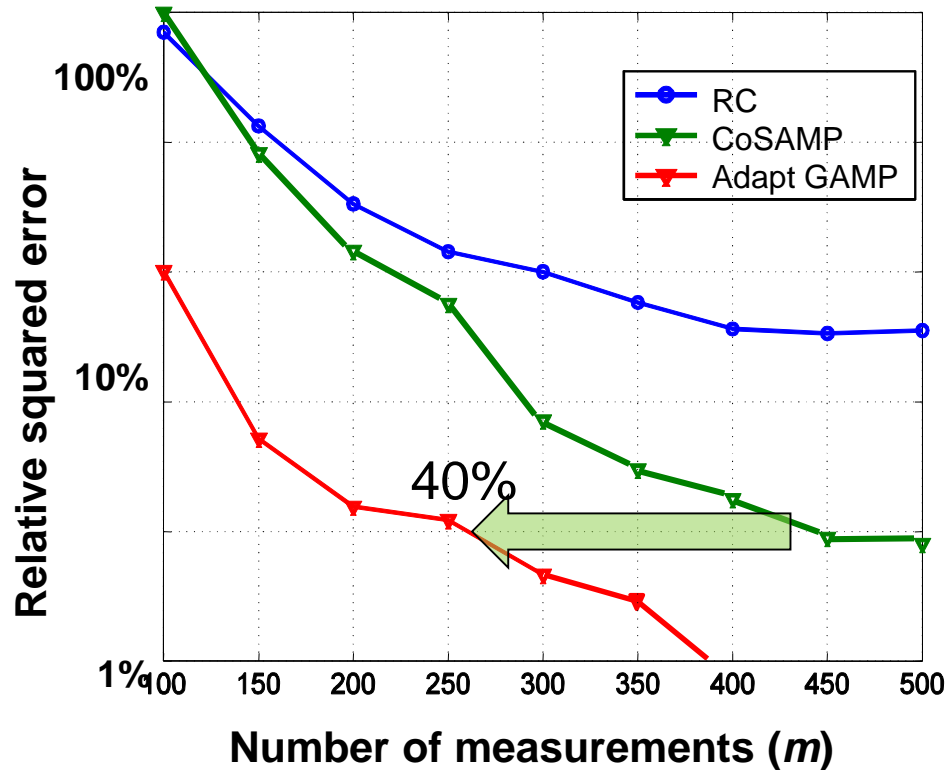
Neural Mapping via Adaptive-GAMP: NeuRAMP



- **Problem:** For neural LNP model:
 - Incorporates sparsity on prior $P_X(\mathbf{x})$
 - $P_{Y|Z}(\mathbf{y}|\mathbf{z}, \theta_z)$ models unknown output nonlinearities
- **Jointly:** Estimate weight vector \mathbf{x}
Learn the nonlinearity
- Computationally fast
- Improved estimates with fewer measurements

Fletcher et al. NIPS (2011)

Simulation: MSE of Synaptic Weight Estimates



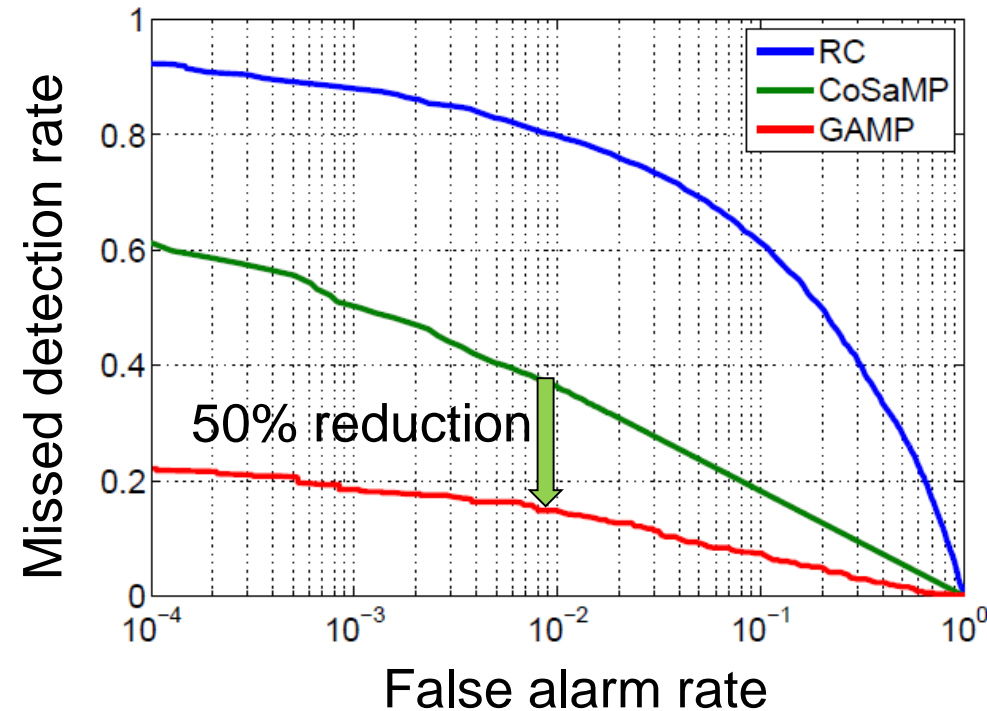
- RC: Reverse Correlation
 - Linear estimation
 - No sparsity
- *CoSaMP: Greedy CS
 - Ignores nonlinearities
- **Adaptive GAMP:
 - Lower MSE
 - Fewer measurements

m = number of trials, random excitation ~ 40
 $n = 500$,
 $k = 30$ or 6% Bernoulli-Gayssuab weights
100 spike windows: 10.4s
Spike rate: 10 spikes/s with .4 second reset
300 trials: ~ 1 hour

*Hu & Chlovskii NIPS 2010

**Fletcher et al, NIPS 2011

Simulation: NeuRAMP Connectivity Detection



■ Factors:

- Models nonlinearities
- Optimal learning
- Incorporates sparsity
- Similar complexity

■ GAMP outperforms RC & CoSaMP

- 75% and 50% lower missed detects

“Simultaneous all optical manipulation and recording...”, AM Packer, Dec 2014 Nature

****“Block sparse” filters: Salamander visual receptive field**

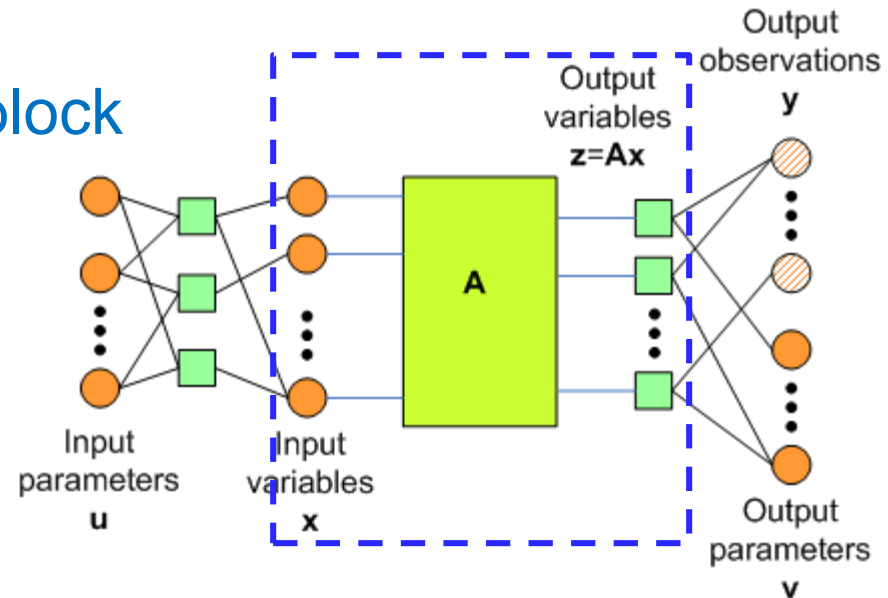


Moving forward: Scalable adaptive block

- Generalized Approximate Message Passing (GAMP)
- Improved neural connectivity detection
 - Data limited
 - Unknown nonlinearities, sparsity levels

- Scalable adaptive GAMP block

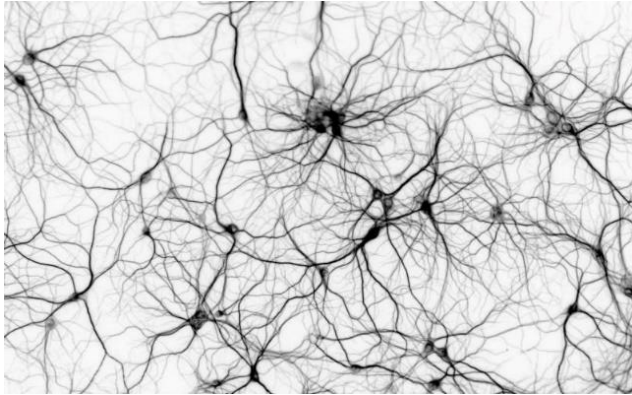
- Linear mixing blocks
- Low complexity: scalable
- Extensible
- Rich input output models
- Adaptive



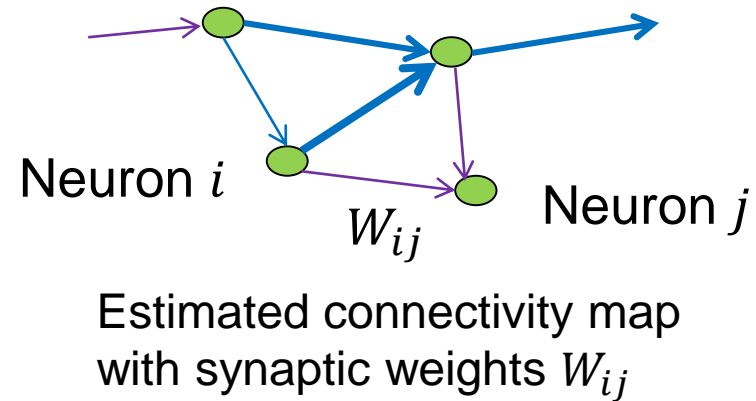
Outline

- Connectivity via multineuronal stimulation
 - Iterative fast adaptive GAMP framework
- Network connectivity via Ca^{2+} imaging
 - Large scale *in vivo* layers
 - Remarkable spatial resolution
 - Indirect, temporally smoothed
 - Network model - captures dynamics
 - Scalable accurate EM algorithm
- Receptive field of retinal ganglion cells
 - Space-time salamander response to stimuli
 - Improved identification with limited data

Inference of Network Connectivity



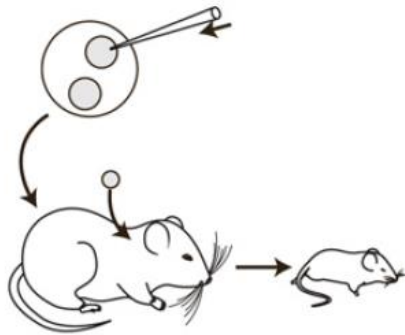
Network of neurons:
unknown synaptic connectivity



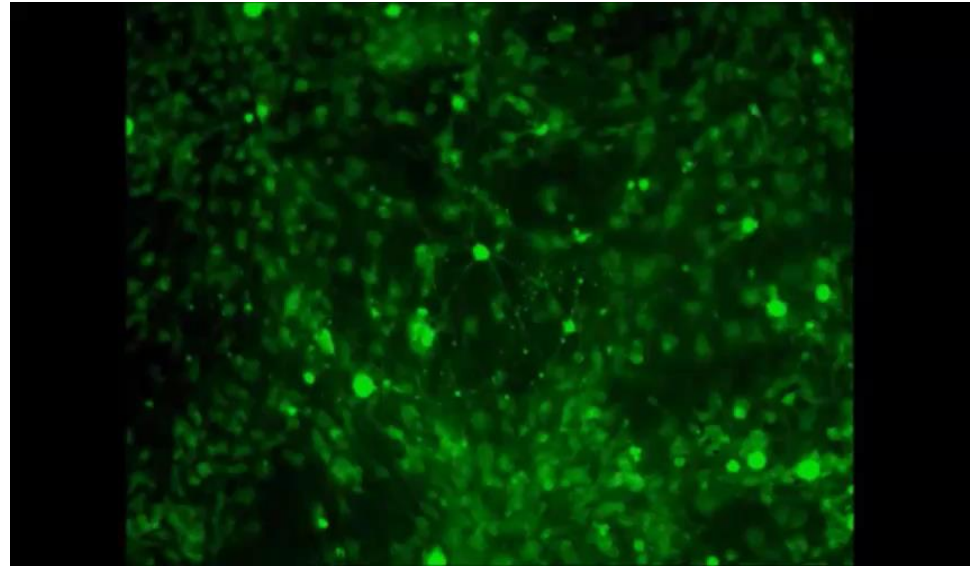
Estimated connectivity map
with synaptic weights W_{ij}

New Technologies: Calcium Imaging

- Fluorescent Ca^{2+} indicators
 - Genetically encoded
 - Chemical dyes
- Spiking: Ca^{2+} influx
- Large populations in parallel



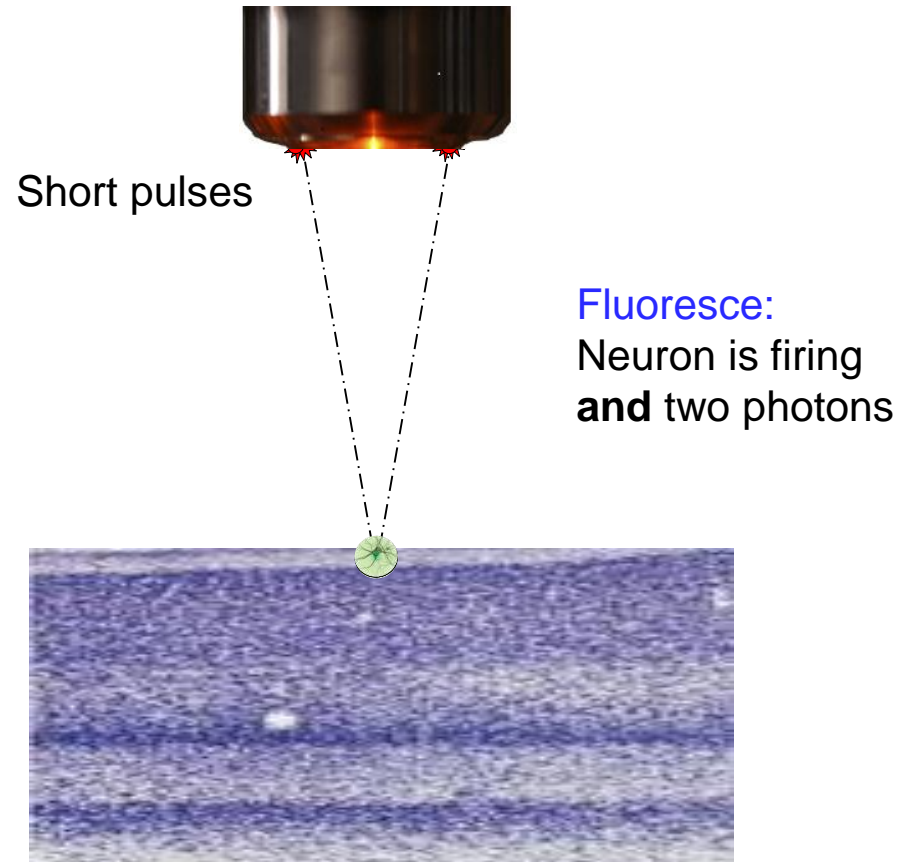
Transgenic mice



Mei Zhang 2009

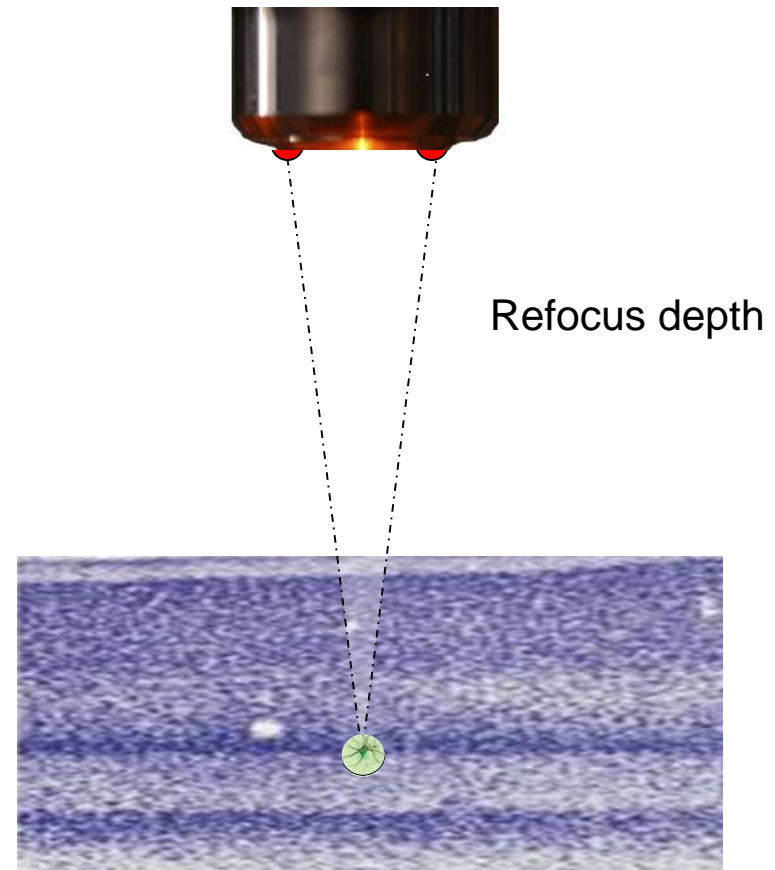
Two photon Ca^{2+} imaging: depth acquisition

- Fluorescence & spiking:
 - Ca^{2+} influx
- Two-photon imaging
 - Raster scan
 - Depth acquisition
- Spatial resolution
 - Image into the cortex



Two photon Ca^{2+} imaging: depth acquisition

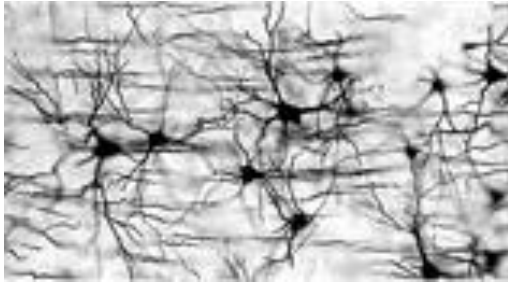
- Fluorescence & spiking:
 - Ca^{2+} influx
- Two-photon imaging
 - Raster scan
 - Depth acquisition
- Spatial resolution
 - Image into the cortex



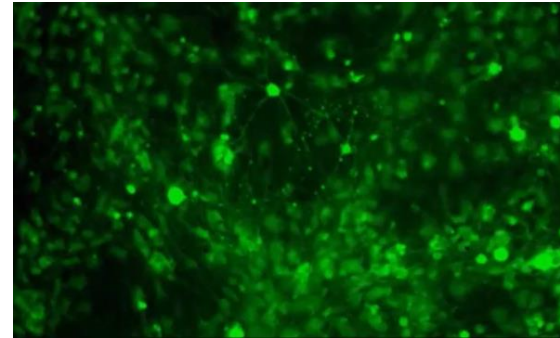
Cortical layers

Cortex ~ 2-4 mm

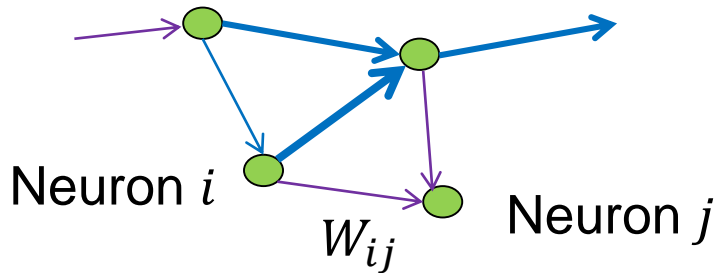
Calcium Imaging: Connectivity Detection Problem



Network of neurons:
unknown synaptic connectivity



Ca²⁺ fluorescence movie



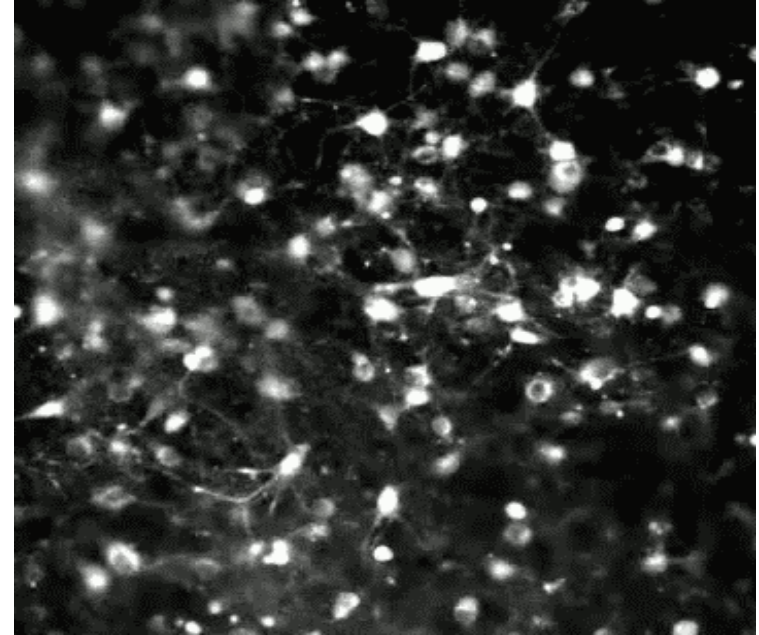
Estimated connectivity map
with synaptic weights W_{ij}



Ca²⁺ fluorescence image
MPI 2012

Ca²⁺ Imaging: Strengths

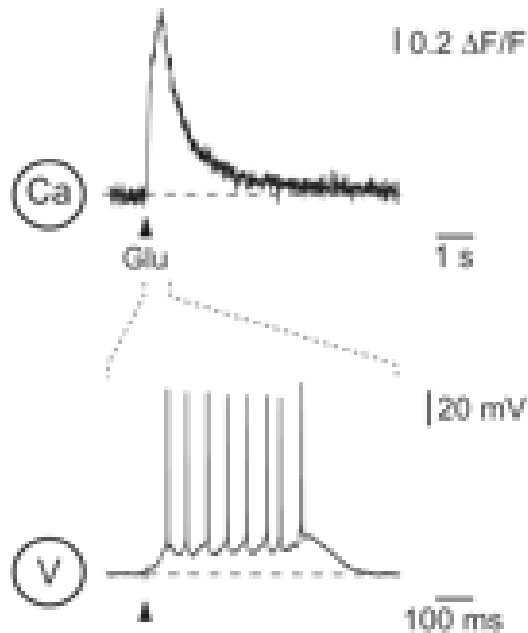
- Parallel measurements
($\sim 10^3$ neurons)
- *In vivo* or *in vitro*
- High spatial resolution
(neuronal level, sub- μm)
- Image below surface



M. Kuykendal and G. Givanasen, Georgia Tech

Challenges with Ca²⁺ Imaging

Calcium Fluorescence



[Stociek et al. (2003)]

- Indirect fluorescence traces
- Nonlinear dynamics
- Large data sets
- Exogenous inputs
- Heavy temporal blurring
 - Ca²⁺ long decay: $\tau_c \sim 0.5$ s **
- Low frame rate
 - Frames: 10-100 ms*
 - Interneuron dynamics: 1-3 ms
- Need **super-resolution**

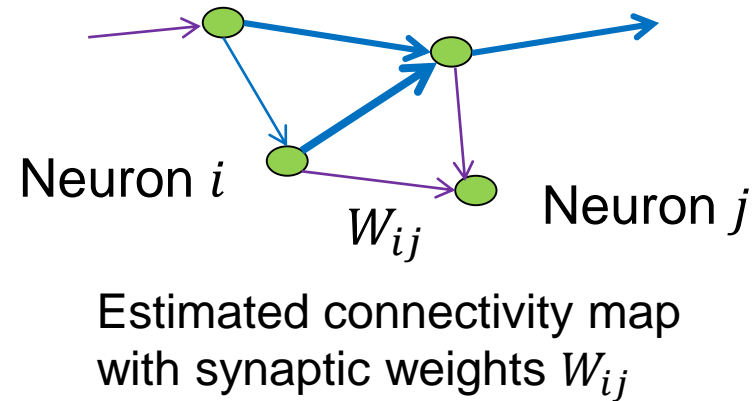
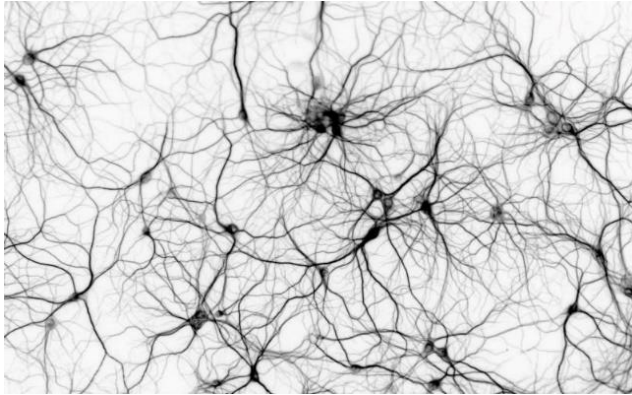
**GCaMP5, newer indicators faster

Glutamate induced spiking

NIPS 2013 & NIPS 2015 workshops:

"Statistical Methods For Understanding Neural Systems"

Network Inference: Causality Crucial

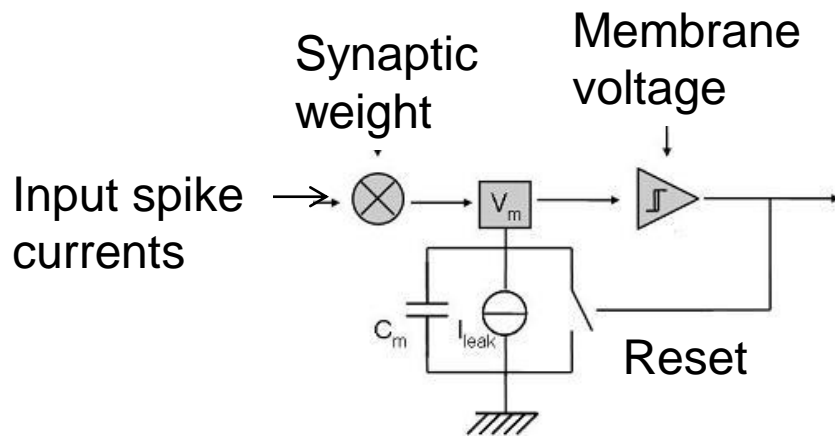


Network of neurons:
unknown synaptic connectivity

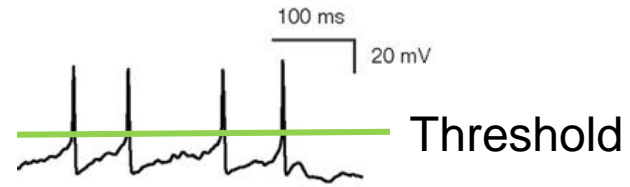
Estimated connectivity map
with synaptic weights W_{ij}

Elements of the network : time-varying electrochemical devices

Neuronal Model: Integrate and Fire



Spike emission



- Electrochemical dynamic model:

- $V_i(t)$ = neuron i potential $I_{ij}(t)$ = current j^{th} to i^{th} neuron

- Integrate phase: Potential builds: $V_i(t) \leq V_{th}$

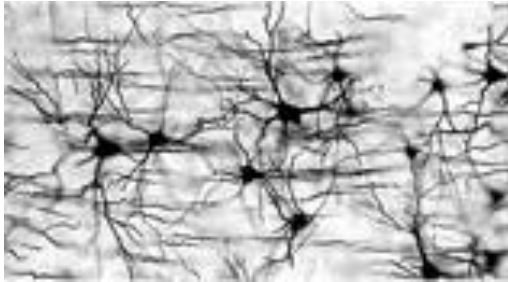
$$RC_m \frac{dV_i(t)}{dt} = \underbrace{-V(t)}_{\text{leakage}} + \underbrace{\sum_{j=1}^N R_{ij} I_{ij}(t)}_{\text{Incoming current}}$$

Charge increase

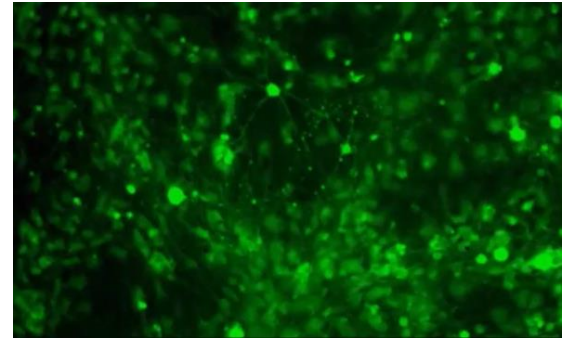
- Fire: discharges spike, reset: $V_i(t) = V_{th} \Rightarrow V_i(t^+) = V_{reset}$

[Lapicque (1907)]

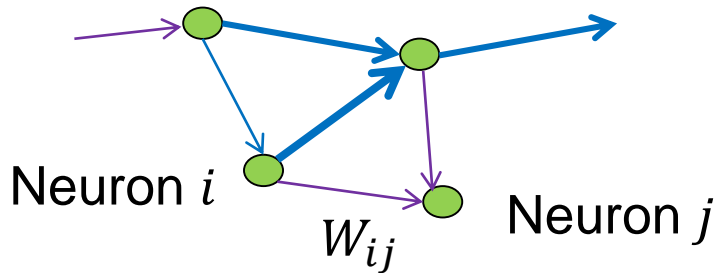
Calcium Imaging: Connectivity Detection Problem



Network of neurons:
unknown synaptic connectivity



Ca²⁺ fluorescence movie



Estimated connectivity map
with synaptic weights W_{ij}



Ca²⁺ fluorescence image
MPI 2012

Each Neuron: Discrete-Time Neural Model

- Voltage: integrate and fire:

$$v_i^{k+1} = (1 - \alpha)v_i^k + \sum_{j=1}^n W_{ij} s_j^k + d_{v,i}^k \quad [\text{Integrate}]$$

$$\text{if } v_i^{k+1} \geq \mu \Rightarrow s_i^k = 1, v_i^{k+1} = 0, \quad [\text{spike \& reset}]$$

$$\text{else } v_i^{k+1} < \mu \Rightarrow s_i^k = 0 \quad [\text{no spike}]$$

Calcium fluorescence

$$z_i^{k+1} = (1 - \beta)z_i^k + s_i^k + d_{z,i}^k \quad [z: \text{Ca}^{2+}]$$

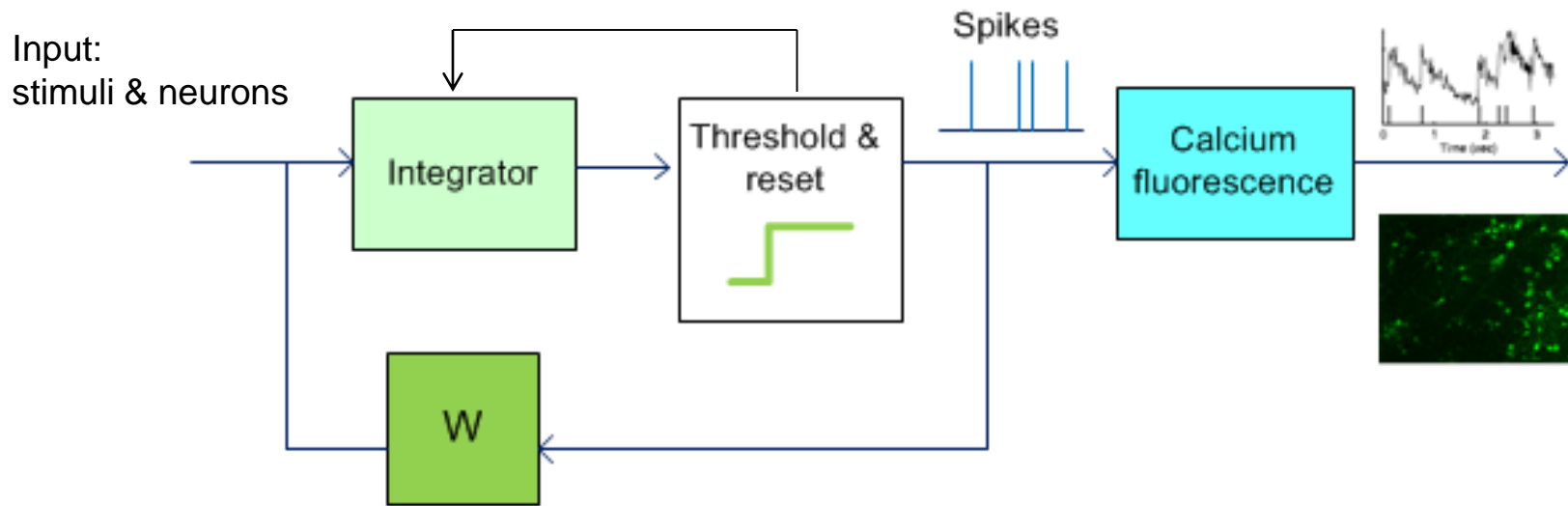
$$y_i^k = a z_i^k + d_{y,i}^k \quad [y: \text{fluorescence}]$$

W_{ij} = "weight" = integrated voltage change from spike current

- Ca^{2+} fluorescence: dynamical system also
- Nonlinear state space
- Need: connectivity, spike times, voltages, calcium...

Mischenko, Vogelstein, Paninski (2010), Yasuda(2004), Vogelstein et al (2010)
Fletcher et al, COSYNE 2014, NIPS 2014

Summary: System



$$v^{k+1} = (1 - \alpha)v^k + Ws^k + d_x^k \quad \leftarrow \text{Membrane voltage integration}$$

$$s^k = 1, v^k = 0 \text{ when } v^k > \mu \quad \leftarrow \text{Spike and reset}$$

$$z^{k+1} = (1 - \beta)z^k + d_z^k + s^k \quad \leftarrow \text{Bound Ca}^{2+} \text{ concentration}$$

$$y^k = a z^k + d_y^k \quad \leftarrow \text{Fluorescence}$$

Maximum Likelihood Estimation of Connectivity

- ML estimate:

$$\hat{W} = \arg \max_W \log p(y|W) - \lambda \|W\|_1$$

Observations

Desired parameters
e.g. connectivity matrix

- Can add regularization term to impose sparsity

Expectation Maximization Algorithm

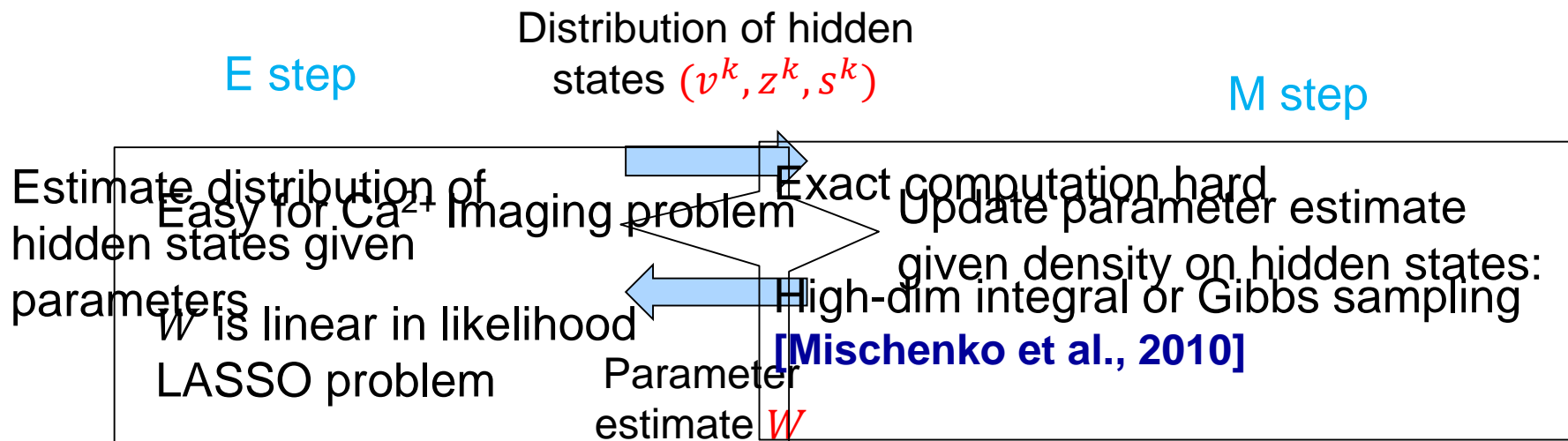
$$\mathbf{v}^{k+1} = (1 - \alpha)\mathbf{v}^k + \mathbf{W}\mathbf{s}^k + \mathbf{d}_x^k$$

$$\mathbf{s}^k = 1, \mathbf{v}^k = 0 \text{ when } \mathbf{v}^k > \mu$$

$$\mathbf{z}^{k+1} = (1 - \beta)\mathbf{z}^k + \mathbf{d}_z^k + \mathbf{s}^k$$

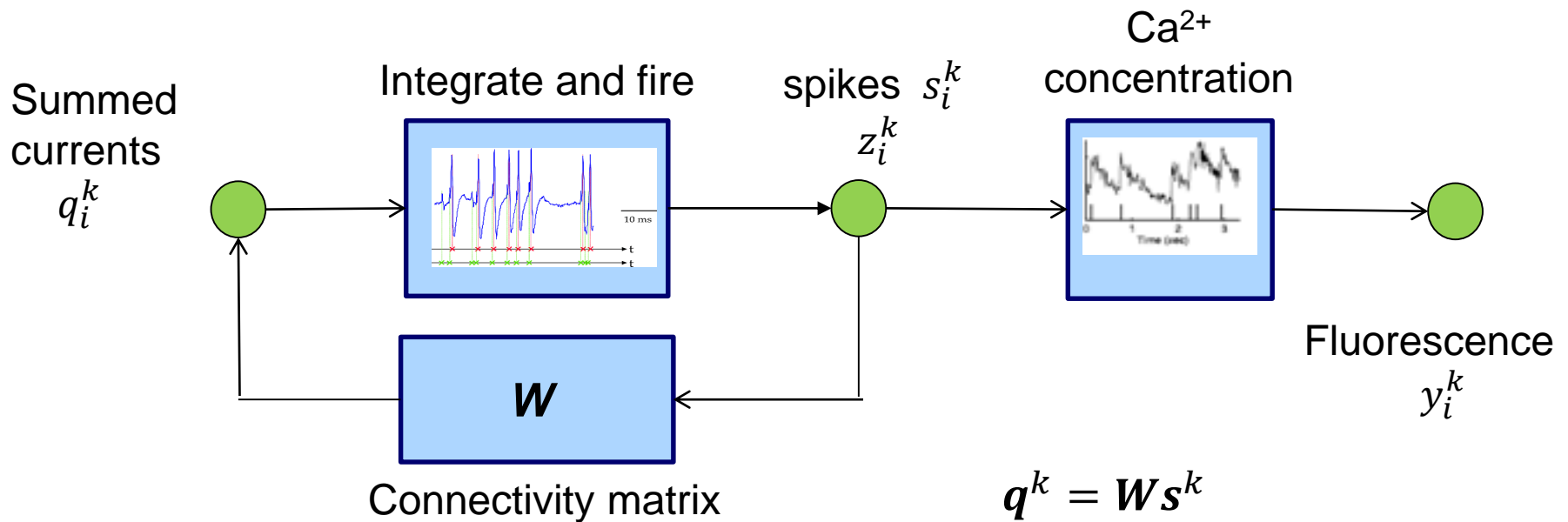
$$\mathbf{y}^k = \mathbf{a}\mathbf{z}^k + \mathbf{d}_y^k$$

- Want regularized ML:
 $\hat{W} = \arg \max_W \log p(y|W) - \lambda \|W\|_1$
- **Problem:** Hidden states $(\mathbf{v}^k, \mathbf{z}^k, \mathbf{s}^k)$
- Use EM iterations



Decoupling for the E-Step

- **Want:** State estimates for nonlinear system
 - High dimensional : N Neurons, 3N states
- **Key insight:** System decouples: scalar iterations $q^k = Ws^k$



Decoupling for the E-Step

N scalar 1-dim systems

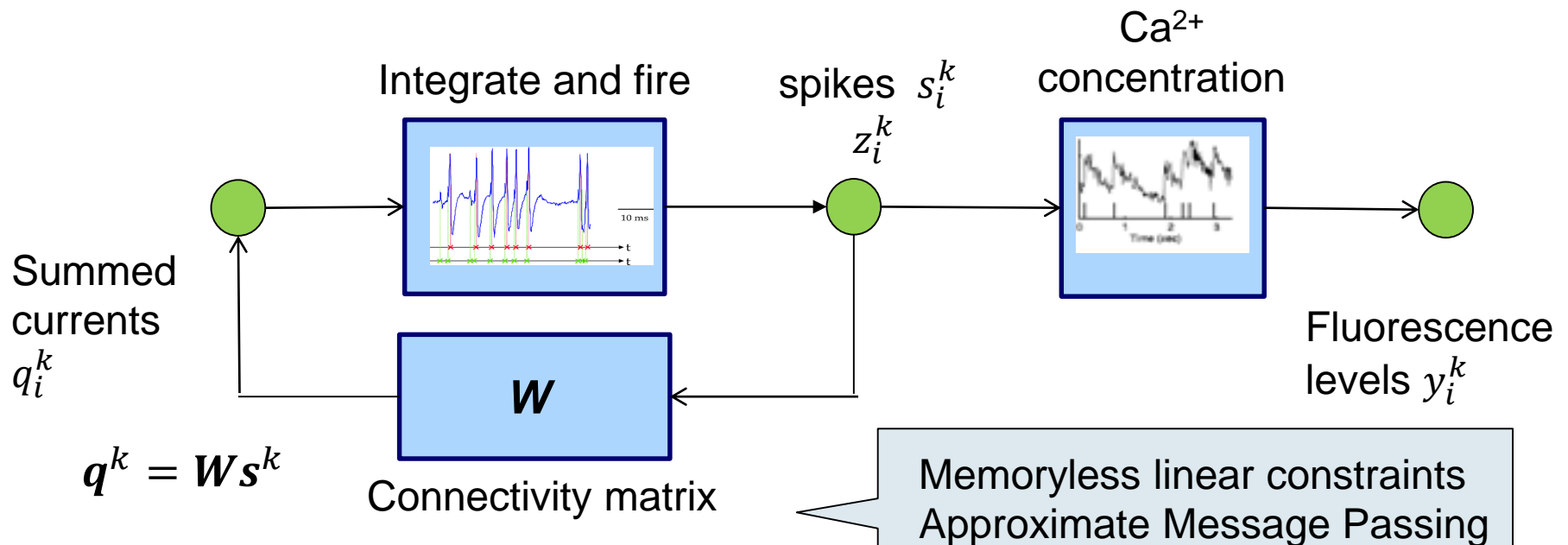
$$v_i^{k+1} = \begin{cases} (1 - \alpha)v_i^k + q_i^k & v_i^k \leq \mu \\ 0 & v_i^k > \mu \end{cases}$$

$$s_i^k = 1 \quad \text{when} \quad v_i^k \geq \mu$$

N scalar 1-dim systems

$$z_i^{k+1} = (1 - \beta)z_i^k + s_i^k$$

$$y_i^k = az_i^k + d_i^k$$



Memoryless linear constraints
Approximate Message Passing

Decoupling for the E-Step

N scalar Forward Backward

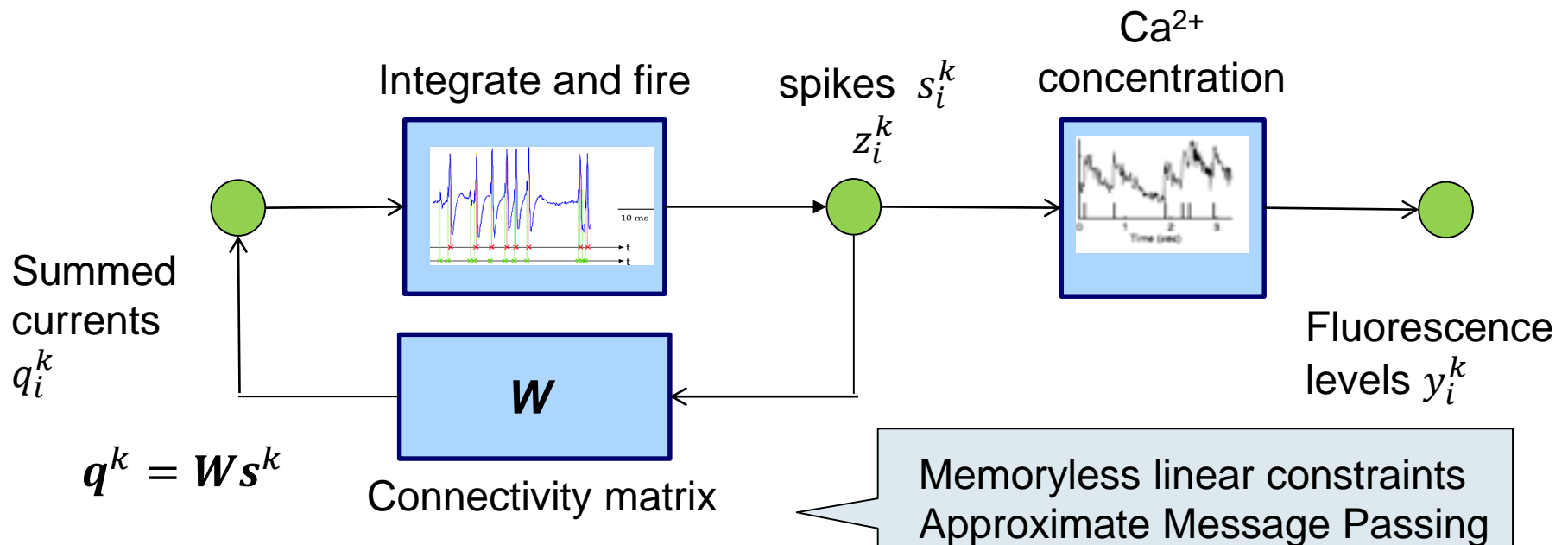
$$v_i^{k+1} = \begin{cases} (1 - \alpha)v_i^k + q_i^k & v_i^k \leq \mu \\ 0 & v_i^k > \mu \end{cases}$$

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N scalar Forward Backward

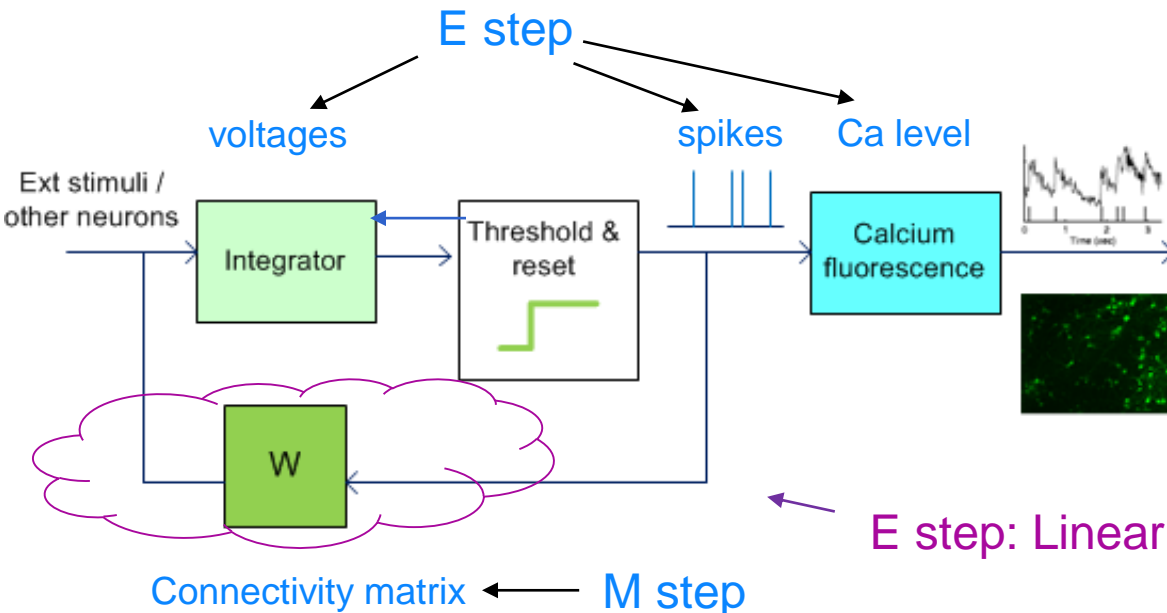
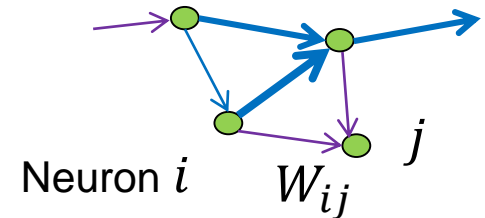
$$z_i^{k+1} = (1 - \beta)z_i^k + s_i^k$$

$$y_i^k = az_i^k + d_i^k$$



EM Overview

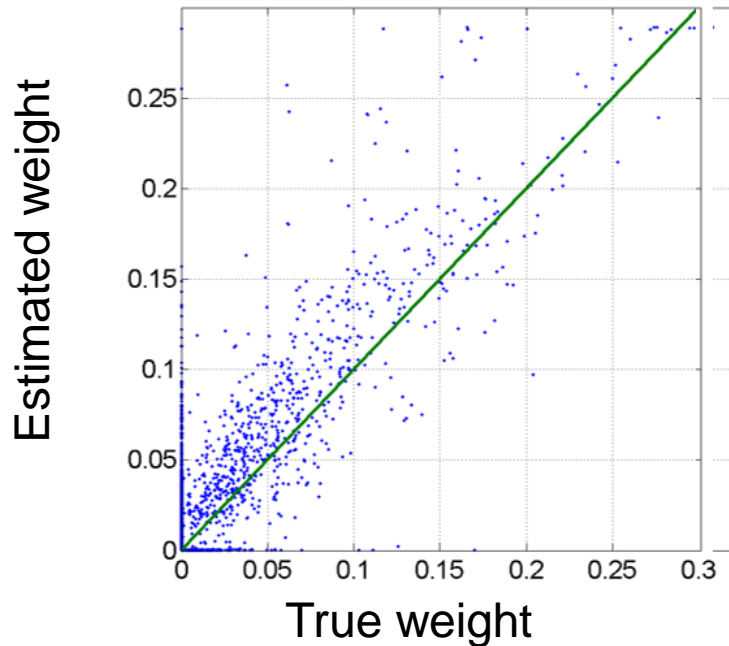
- Fluorescence movie: estimate
 - Connectivity matrix W , spike times, voltages
- Scalable block
 - High-dimensional, nonlinear dynamical system



E step: Linear Mixing AMP block

Simulation Results: Accuracy of the Weights

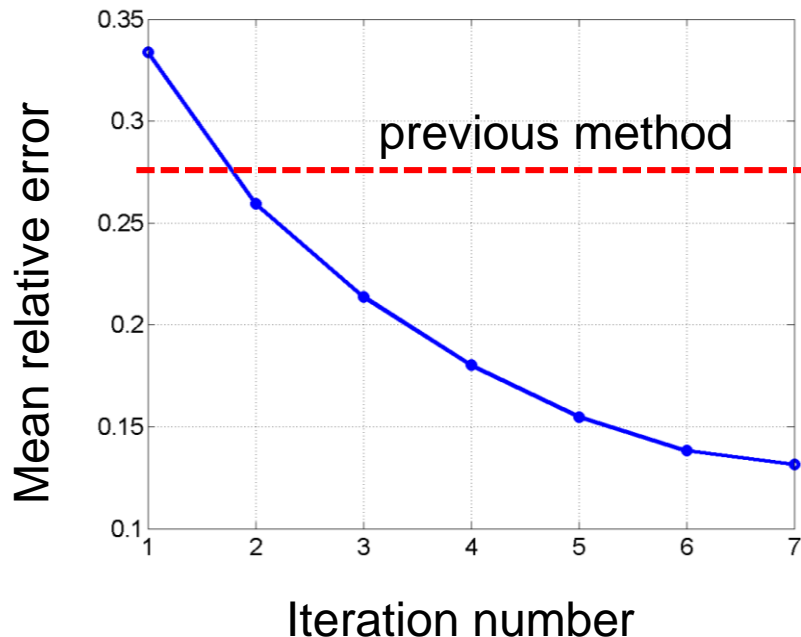
Accurate estimation



- Neuron model [Sayer (1990)]
 - 100 guinea pig cortical neurons
 - Synchronized bursting: 10 spikes/s
 - 10% sparse random connectivity
 - 20 ms integrate-fire τ_c
 - 2 ms inter-neuron conduction time
 - 1 ms time step
- Ca^{2+} imaging model
 - 100 frames/s, 100 s trials
 - 10000 $\mathcal{F}l$ values per neuron
 - 500 ms Ca^{2+} τ_c
 - Fluorescence SNR = 20 dB

Accuracy of the Weights

Fast convergence



$$\bullet \text{Relative MSE} = \frac{E(W_{ij} - \widehat{W}_{ij})^2}{E(W_{ij})^2} = 0.12$$

- Neuron model [Sayer (1990)]
 - Guinea pig cortical column
 - 100 neurons
 - 10% sparse random connectivity
 - Synchronized bursting: 10 spikes/s
 - 20 ms integrate-fire τ_c
 - 2 ms inter-neuron conduction
 - 1 ms time step
- Data Collection
 - 100 frames/s, 100 s trials
 - 10000 $\mathcal{F}l$ samples per neuron
 - Ca^{2+} $\tau_c = 500$ ms, $\mathcal{F}l$ SNR = 20 dB

- Previous work MSE=0.28, same parameters

Mischenko et al. (2010)

- Accurate, low complexity $O(N)$ per iteration, instead of $O(*^N)$

Fletcher et al NIPS '14

Network Connectivity Summary

- Network analysis from Ca^{2+} imaging
 - Attack temporal resolution issues for neural dynamics
 - Scalable EM algorithm
- Rich, flexible modeling framework
 - Incorporates nonlinearities, indirect measurements, dynamics
- Computationally scalable solution
 - Linear in number of measurements
- Demonstrated performance
 - Outperforms existing techniques
 - Allows more biologically plausible model with feedback

<http://gampmatlab.sourceforge.net/>

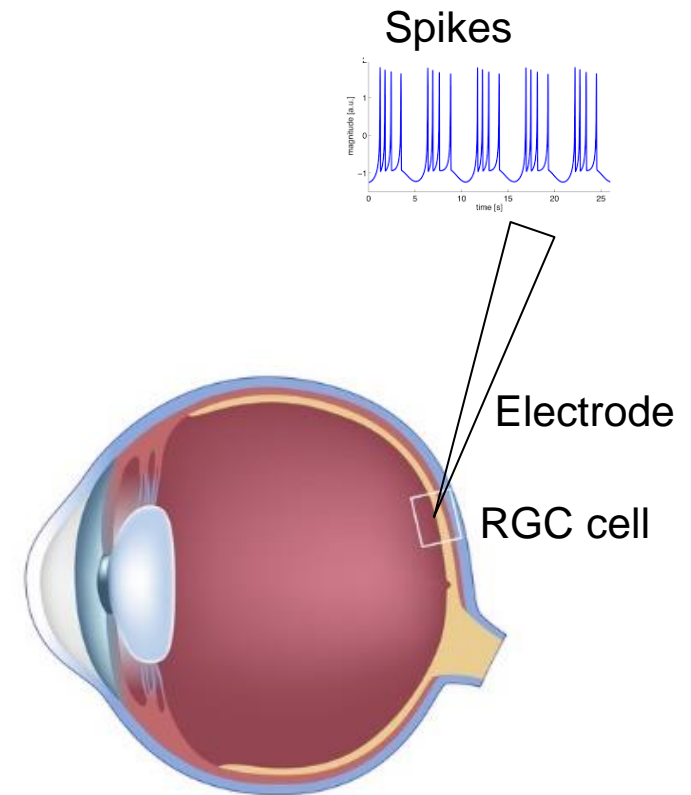
Phil Schniter!!!

Outline

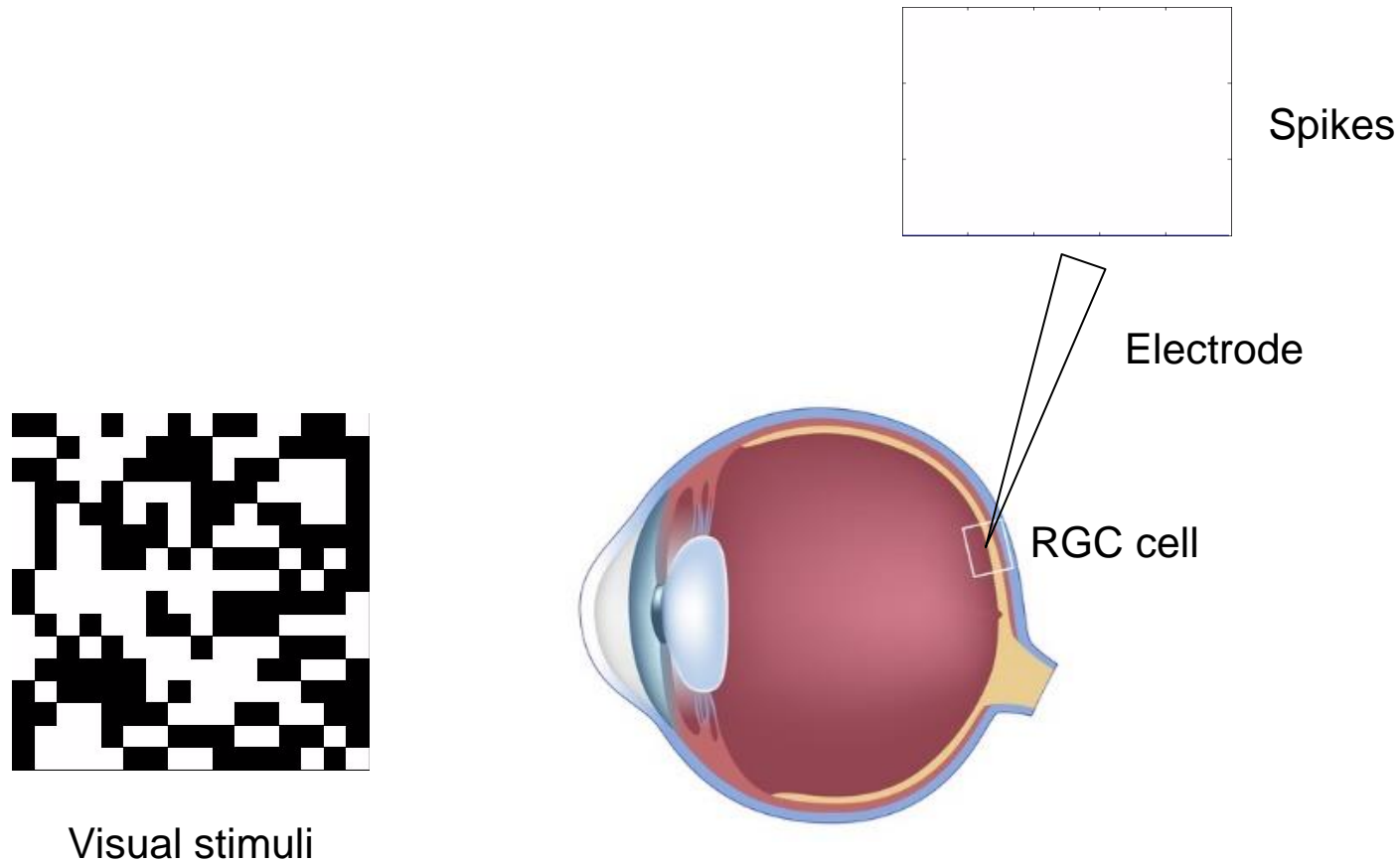
- Connectivity via multineuronal stimulation
 - Iterative fast adaptive GAMP framework
- Network connectivity via Ca^{2+} imaging
 - Network model - captures dynamics
 - Graphical models
 - Scalable accurate based algorithm
- Receptive field of retinal ganglion cells
 - Space-time salamander response to stimuli
 - Improved identification with limited data

Receptive Field Identification

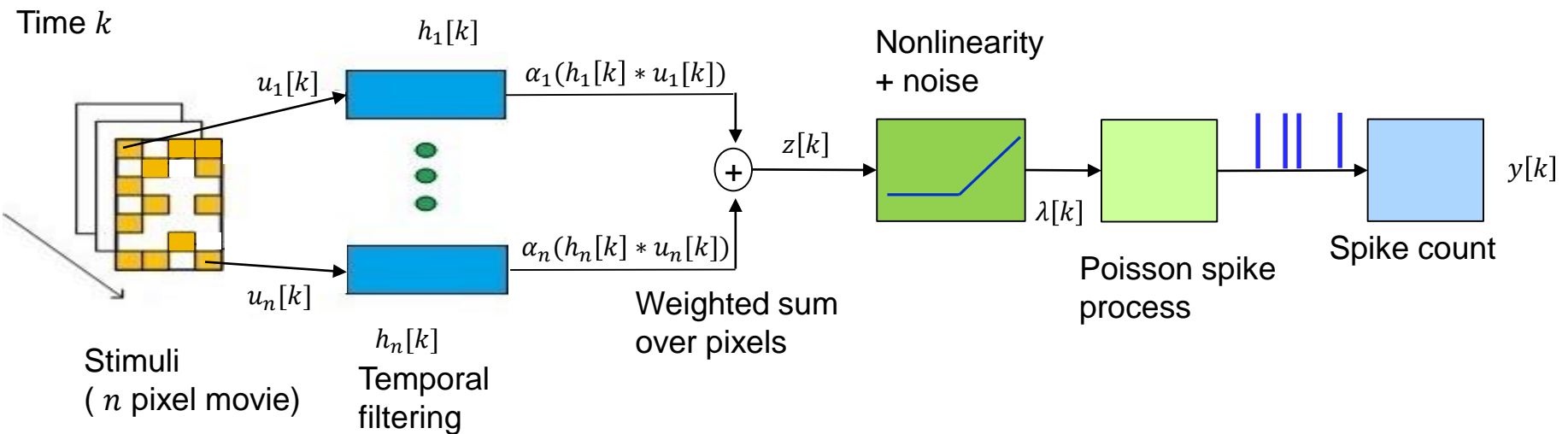
- **Retinal ganglion cell (RGC)**
Sensitive to light in its field of view,
or receptive field
- Tuned to some local features
in time and space (curve, edge, etc)
- Response estimation of RGCs:
 - Expose retina to image
 - Measure response via electrode
 - Fit model
- Challenge: Model is often nonlinear



Salamander Receptive Field Identification



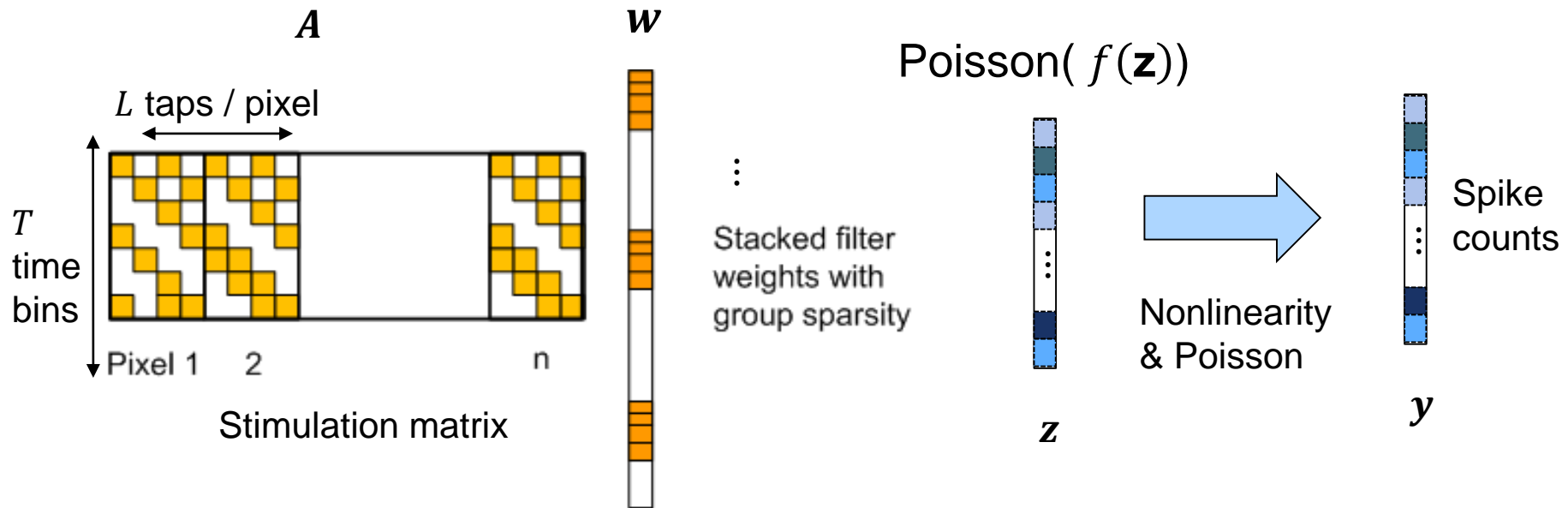
Retinal Ganglion Cell LNP Model



- Linear-Nonlinear Poisson model. Given stimuli $u_i[k]$
 - Filtering over time and space: $z[k] = \sum_{i=1}^n \alpha_i(h_i[k] * u_i[k])$
 - Nonlinear phase: $y[k] = \text{Poisson}(f(z[k] + d[k]))$
- Identification problem: Given stimuli $u_i[k]$ and spike counts $y[k]$
 - Estimate weighted filters $w_i[k] = \alpha_i h_i[k]$,
Describes space-time response of neuron to pixel i
 - Each $h_i[k]$ is an L tap filter
 - Estimate nonlinearity $f(\cdot)$

Fletcher et al. NIPS 2012

Structured Matrix View of Dynamic LNP



- LNP model: cascade of linear and nonlinear system
 - A rows: n pixel values at L delays ($L =$ filter taps)
- Weights have a **group sparse** constraint:
 - RGC sensitive to small image region (spatial sparsity)
 - Coefficients of filter of one stimuli are on or off together

Classical Methods and their Limitations

■ Linear methods:

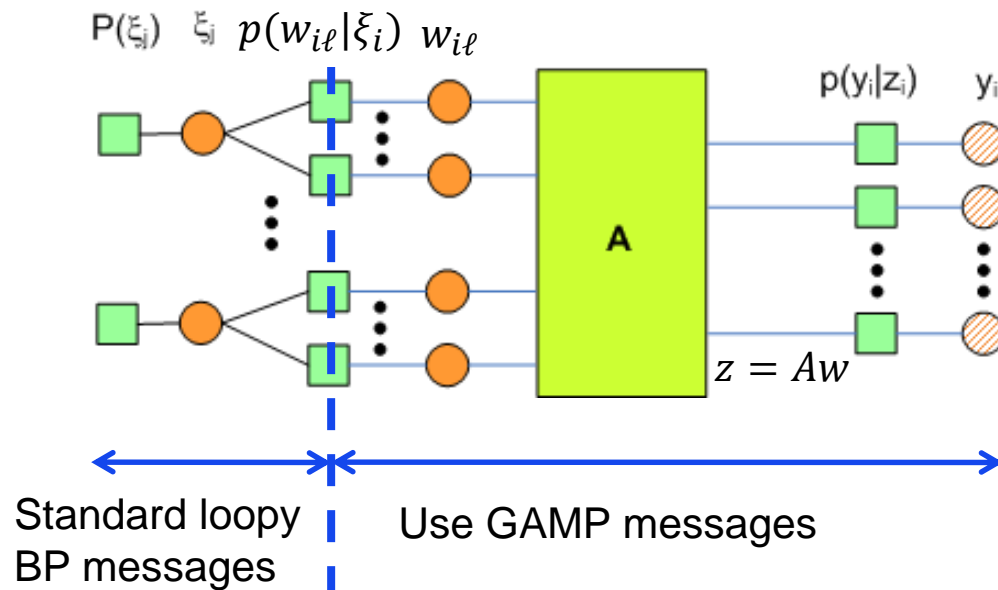
- Matched filter: $\hat{\mathbf{w}} = \frac{1}{n} \mathbf{A}^T \mathbf{y}$ (also called STA)
- Linear MMSE: $\hat{\mathbf{w}} = (\mathbf{A}^T \mathbf{A} + \sigma^2 \mathbf{I})^{-1} \mathbf{A}^T \mathbf{y}$ (also called RC)
- Also, linear least squares / zero forcing
- Simple but does not exploit sparsity

■ Compressed sensing methods

$$\hat{\mathbf{w}} = \arg \min_{\mathbf{w}} \|\mathbf{y} - \mathbf{A}\mathbf{w}\|_2^2 + \lambda \|\mathbf{w}\|_1$$

- Exploit sparsity of \mathbf{w}
- Many methods: LASSO, OMP, CoSAMP,...
- Could also incorporate group sparsity via group Lasso
- But, does not account for output nonlinearities

Hybrid Algorithm for Structured Input: GAMP

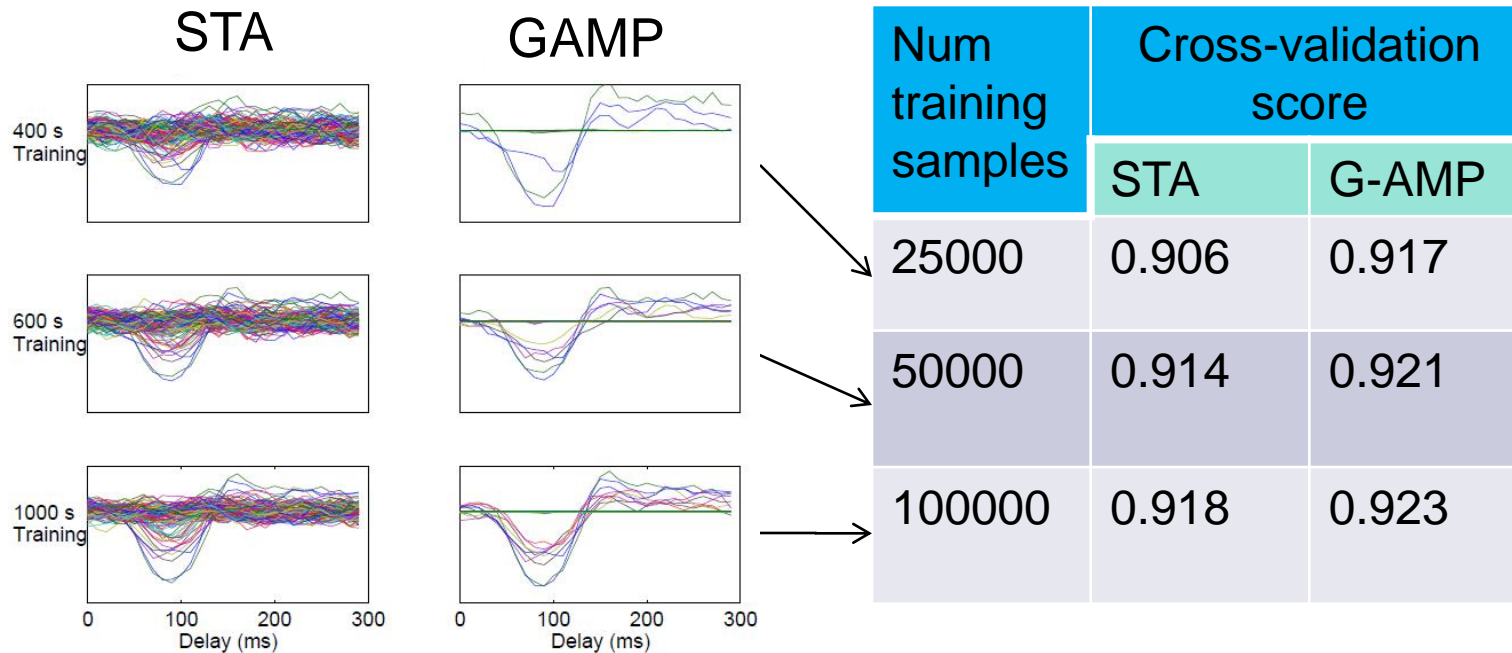


- Introduce binary variables ξ_j to correlate sparsity over time
- Apply GAMP in a “turbo” manner with loopy BP
- Low complexity: each ξ_j is binary
- More general than group OMP and group lasso;
 - Similar complexity,
 - Better performance [Rangan, Fletcher, Goyal & Schniter ‘12]

Receptive Field Computation Considerations

- Problem size: 3630 variables
 - 11 x 11 pixels, 30 taps per pixel
 - 190,000 measurements (~30 min at 10 ms sampling)
 - Structured A matrix is 190,000 by 3630
- AGAMP iteration cost: multiplying by A & A^*
 - Exploit block Toeplitz structure and entries are 0-1
- For larger problems, algorithm is parallelizable
 - Graphical methods: inherent decomposable
 - Parallelize multiplications across rows/columns of A
- Cannot theoretically guarantee convergence
 - Demonstrate performance experimentally

Experimental Results: Cross-Validation



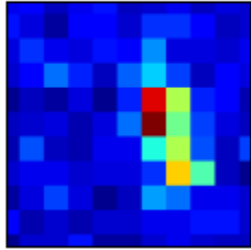
- Validated on data used in training (190000 total samples)
- Cross-validation score = Geometric mean of likelihood of spike rate
- GAMP: same error, 25000 samples versus 100,000

Data: Anthony Leonardo
Janelia Farm

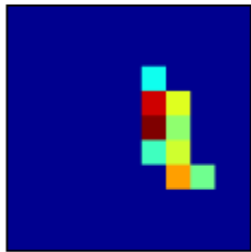


Salamander Retinal Response (Spatial)

Non-sparse LNP w/ STA



Sparse LNP w/ GAMP



Spatial receptive field estimates for 11x11 pixel area



0

1

(normalized)

- **Spatial receptive field:**

- Plot estimated 11x11 response magnitudes.
Color = 30-tap filter magnitude for each pixel

- Standard STA estimate shows noisy (spurious) responses outside

- GAMP method removes noise
Shows only a response in a small area

Filter response 11 x 11 pixels for salamander RGCs
Data from Anthony Leonardo, Janelia Farm



AMP++ methods: Applications in Imaging

- Hybrid-AMP can incorporate complex structure
 - Incorporate dependencies between wavelet coefficients
 - Hierarchical models, etc



<http://gampmatlab.sourceforge.net/>

Algorithm	NMSE (dB)	Comp time (secs)
MHT+IRWL1	-14.37	363
CoSAMP	-16.90	25
SPLG1	-18.06	536
MCMC	-20.10	742
Turbo-GM	-20.74	51

Lowest MSE and almost fastest computation

[Som, Schniter, 2011]

Thank you



Much thanks! To: Fritz Somer, Surya Ganguli, Eero Simoncelli, Eftychios Pnevmatikakis, Matthias Bethge, Anthony Leonardo, Evan Lyall, students of Deisseroth & Adesnik labs, Liam Paninski, Mayur Mudigonda, Jascha Sohl-Dickstein...

■ Funding: NSF CAREER, ONR, AFOSR, Qualcomm



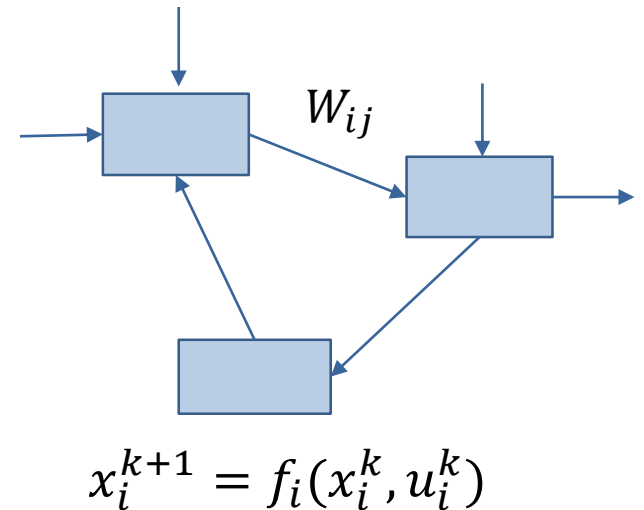
Moving forward

- Better account for exogenous effects
 - Correlations across neurons of interest**
 - Larger areas via “shotgun” techniques [[Pnevmatikakis et al. 2013](#)]
- Data sets in collaboration
 - Paninski Lab, Columbia; Tolias Lab, Baylor, Allen Institute
 - Validate methods: first cultured without exogenous
- Model new Ca^{2+} indicators:
 - GCaMP6f (2013): faster decay, rise time = 50-75 ms,
 - Nonlinear fluorescence model
- Theory
 - Convergence issues of GAMP via new ADMM GAMP
 - Networks of low-dimensional, nonlinear dynamical blocks
 - Provable results for structured non-iid, structured A

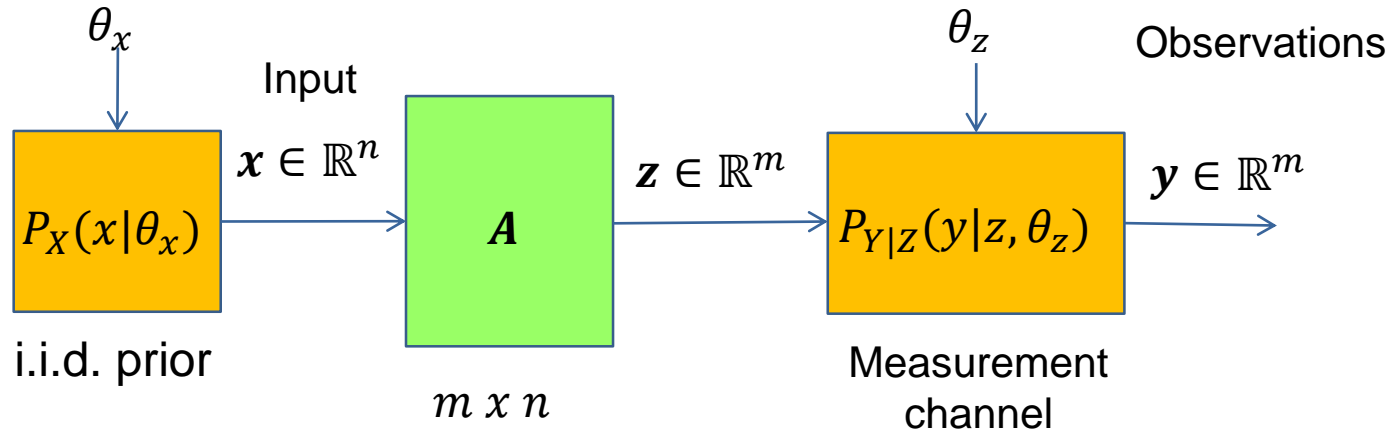
Future work : dynamical networks

High dimensional inference for dynamical systems

- *Generalized linear dynamical networks:*
 - Underlying low-dimensional, nonlinear dynamical blocks
 - Linear memoryless constraints, graphical models
- Many phenomena
 - Neural systems, communication networks, particles, media, ...
 - Extends GLM to include networked dynamics
- Can we extend methods for:
 - Scalable estimation algorithms?
 - Learning connectivity?
 - Provable guarantees?



Joint Estimation and Learning for GLMS



- GLM with unknown parameters θ_x and θ_z
 - Unknown prior, nonlinearities, noise...
- Joint estimation learning problem: Given y and A :
 - Estimate input x and z ,
 - Learn parameters θ_x and θ_z in distribution
 - **Consistent**

Fletcher, Rangan NeuRAMP NIPS 2011

Kamilov, Fletcher, et al NIPS 2012, Trans IT 2014

Future work : broader

High dimensional inference for dynamical systems

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 - Underlying low-dimensional, nonlinear dynamical blocks
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